

Evaluation of Cytotoxicity of Different Concentration of van Tulsi (*Ocimum Gratissimum*) on Roots Tip Cells Onion (*Allium Cepa*)

Praveen Kumar Rana¹, Dr. Dharmshila Kumari²

¹Research Scholar, University Department of Zoology / T.M. Bhagalpur University, Bhagalpur, India

²Professor & Head University Department of Zoology / T.M. Bhagalpur University, Bhagalpur, India

Abstract:

The present study aimed to assess the cytotoxic effects of Van Tulsi (*Ocimum gratissimum*) leaf extract at a 5% concentration on root tip cells of Onion (*Allium cepa*) after exposure periods of 72 hours and 96 hours. Fresh Onion (*Allium cepa*) root tips were exposed to Van Tulsi leaf extract at a concentration of 5% for two different time intervals, 72 hours and 96 hours. Control groups with untreated root tips were also maintained for comparison. The cytotoxicity was evaluated through various cytological parameters, including cell viability, mitotic index- Prophase, Metaphase, Anaphase, Telophase.

The findings revealed a significant decrease in cell viability in the treated groups compared to the control at both 72 hours and 96 hours of exposure. Additionally, the mitotic index was markedly inhibited in root tip cells exposed to Van Tulsi extract. The treated cells exhibited an elevated frequency of chromosomal aberrations and micronuclei, indicating genotoxic effects of the extract.

The results of this study demonstrate that Van Tulsi (*Ocimum gratissimum*) leaf extract at a concentration of 5% exerts cytotoxic effects on root tip cells of Onion (*Allium cepa*). These findings contribute to our understanding of the effects of Van Tulsi leaf extract on plant cells and warrant further investigation into its underlying mechanisms of cytotoxic action. In 72 hrs treatment duration mitotic index slightly increased but at 96 hrs duration mitotic index significantly decreased due to decrease in the proportion of prophase and Metaphase cell population. Thus, leaf extract of Van Tulsi act as mitotic inhibitor on onion root tip cells at higher treatment duration.

Keywords: Mitotic index, *Allium cepa*, Cytotoxic, *Ocimum gratissimum*

INTRODUCTION:

Ocimum gratissimum, the medicinal herbs called as Van Tulsi have been used in folk medicine for million years. Simply, in recent times, scientific study of their effects has flourished. The use of species of the *Ocimum* genus in organic agriculture is described in different works especially in the culture of vegetables due to its bactericidal, antihelminthic [1].

Different parts including the leaves, stem, flowers and roots of Van Tulsi can be used. The leaves and stem are used in Chinese medicine for digestive system disorders, diarrhea and kidney infections, cough, especially chest colds, headache, and various types of fever [2].

Furthermore, over the last decade, medicinal plants and their bioactive compounds have attracted the attention of several researchers because of their usefulness in the management and prevention of life-threatening and chronic diseases such as cancer, diabetes, stroke and arthritis [3], as an alternative therapy for the treatment of psychiatric disorders [4]. This plant possesses numerous pharmacological properties such as aet anti-hyperglycaemic [5], [6], anti-inflammatory [7], [8], anti-diarrhoeal [9], anti-anaemic, hepatoprotective [10], anti-hypertensive [11], antibacterial [12], [13], hypoglycaemic [14], [6] antifungal [15] nematocidal [16], insecticidal [17], antimicrobial [14], fungicidal [18] and anti-oxidative properties [19], [20] as well as exhibits many other pharmacological activities.

The present study was carried out to evaluate the cytotoxic effect of *Van Tulsi (Ocimum gratissimum)* leaf extract on root tip cells of Onion (*Allium cepa*).

MATERIALS AND METHODS:

PREPARATION OF LEAF EXTRACT: The leaves of Van Tulsi (*Ocimum gratissimum*) were washed under running tap water and shade dried for 2 to 3 weeks. After that, dried fresh Tulsi leaves were homogenised by using a grinder to made fine powder so obtained and stored in air tight bottles. 100gram fine powder were dissolved in 1000ml distilled water as a stock solution and left for 48hours. It was then filtered through Whatman No. 1 [21]. Van Tulsi leaf extract 5% dose was prepared by dilution of stock solution [22].

The onion bulb weighing approximately 20-30 grams were purchased from local market and their roots were initially allowed to grow till 1.5 cm in length in normal tap water. The bulb roots were cut after 72hrs and 96hrs and fixed in aceto-alcohol for 24hrs then preserved in 70% ethanol and used as control group.

Another set of onion bulbs (20-30gm) were grown in 5% Van Tulsi leaf extract for 72hrs and 96hrs respectively and used as treated group.



Figure 1: (Growing of Control and Treated Onion root tips).

SLIDE PREPARATION:

After treatment, slides were prepared by Acetocarmine squash preparation [23]. Approximately 4000 cells were randomly analyzed in both control and treated group of onion bulbs. Frequency of Mitotic index and Phases distribution were calculated.

SLIDE SCREENING:

All the slides were examined under light microscope. The mitotic index was calculated for determination of cytotoxicity. Mitotic index (MI) was calculated as the ratio between the number of mitotic cells and the total number of cells scored and expressed as percentage and represented by following formulae [23].

$$MI = \frac{\text{Total number of dividing cells}}{\text{Total number of cells observed}} \times 100$$

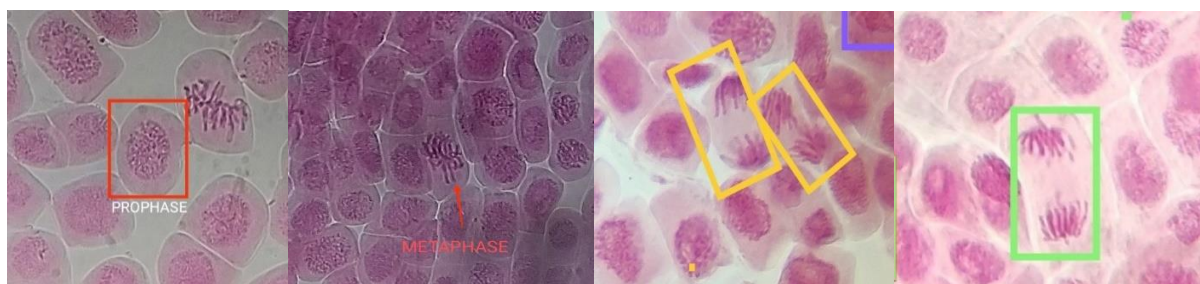
STATISTICAL ANALYSIS:

The data are expressed as Mean ± SE and statistical analysis was performed by using t-test.

Table1: Effect of Van Tulsi (5%) on mitotic index in onion root- tip cells at 72hrs and 96hrs.

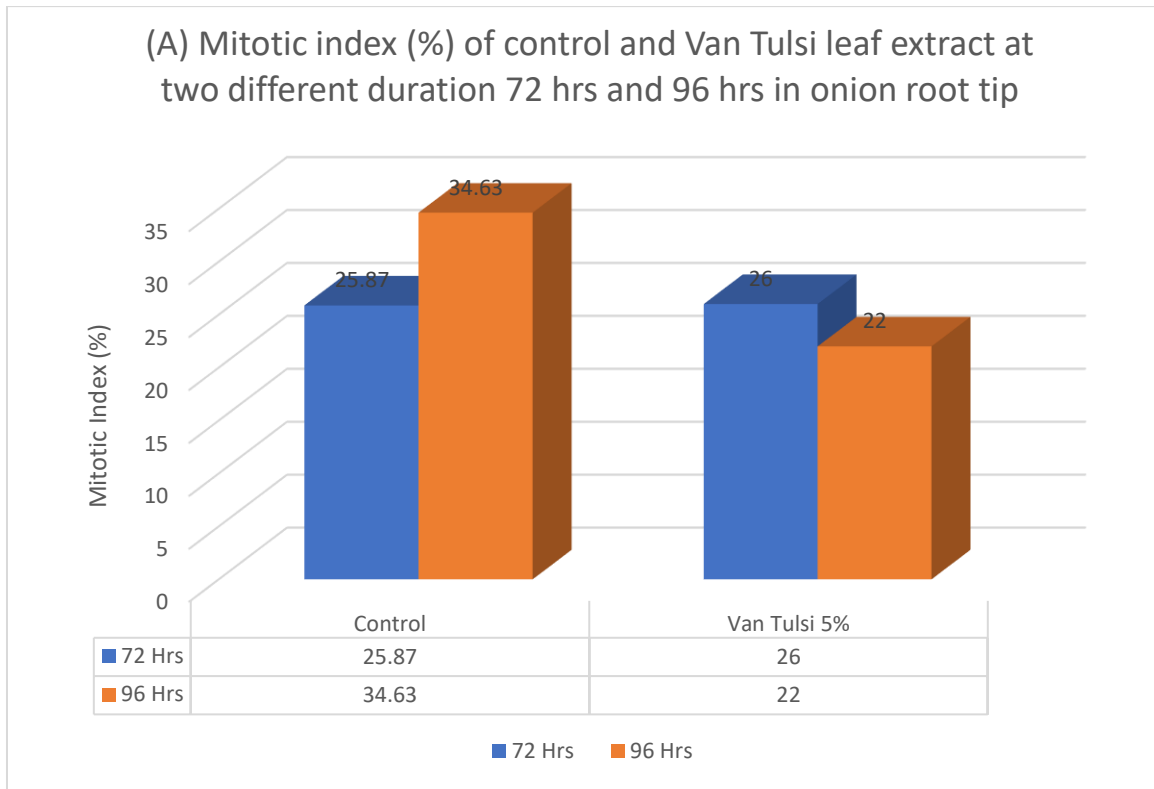
Exp Variat	Durati on (hrs)	Tota l No of Cells Scor ed(N)	Total No of Divi ding cells	Mitotic Index(%±S .E.)	Phase Distribution			
					Prophase (% ± S.E.)	Metaphas e (% ± S.E.)	Anaphase (% ± S.E.)	Telopha se (% ± S.E.)
Contro l	72	3305	855	25.87 ± 0.76	22.72 ± 0.73	1.75 ± 0.23	0.64 ± 0.14	0.76 ± 0.15
V (5%)	72	4853	1262	26.00 ± 0.63	23.10 ± 0.61	1.57 ± 0.18	0.74 ± 0.12	0.60 ± 0.11
Contro l	96	4014	1390	34.63 ± 0.75	29.42 ± 0.72	3.26 ± 0.28	0.97 ± 0.15	0.97 ± 0.15
V (5%)	96	4000	884	22.00 ± 0.65*	18.7 ± 0.62*	2.10 ± 0.23*	0.70 ± 0.13	0.63 ± 0.13

*- Indicate significant difference with control

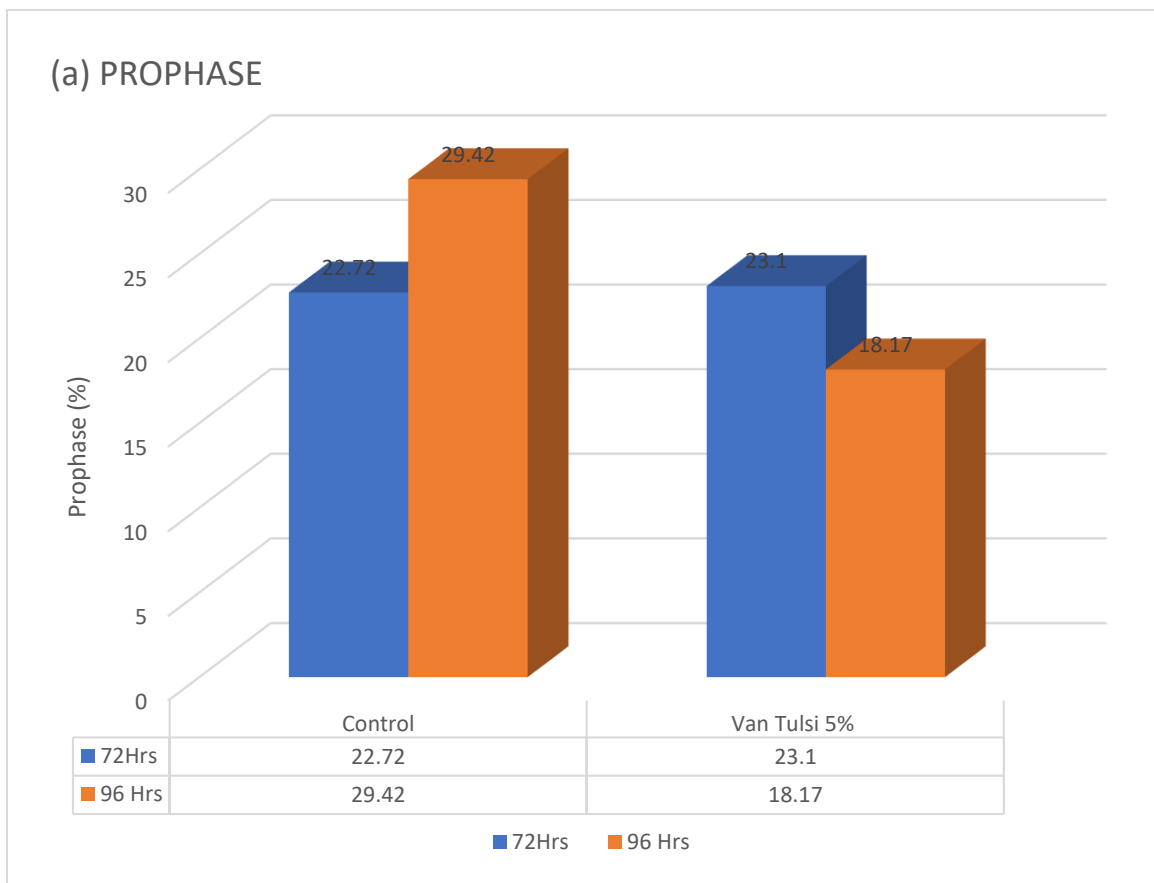


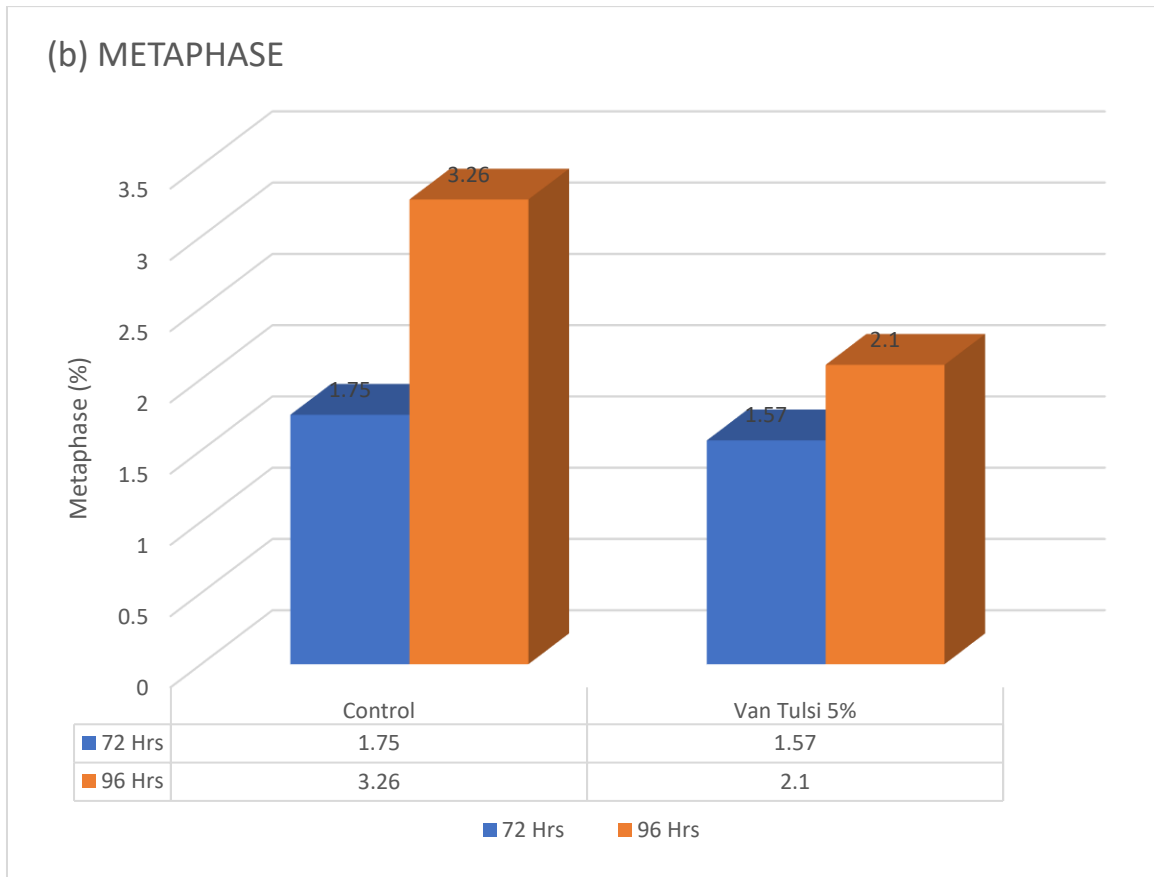
Prophase Metaphase Anaphase Telophase

Figure 2: Different stages of Mitotic division in Onion Root Tip cells.

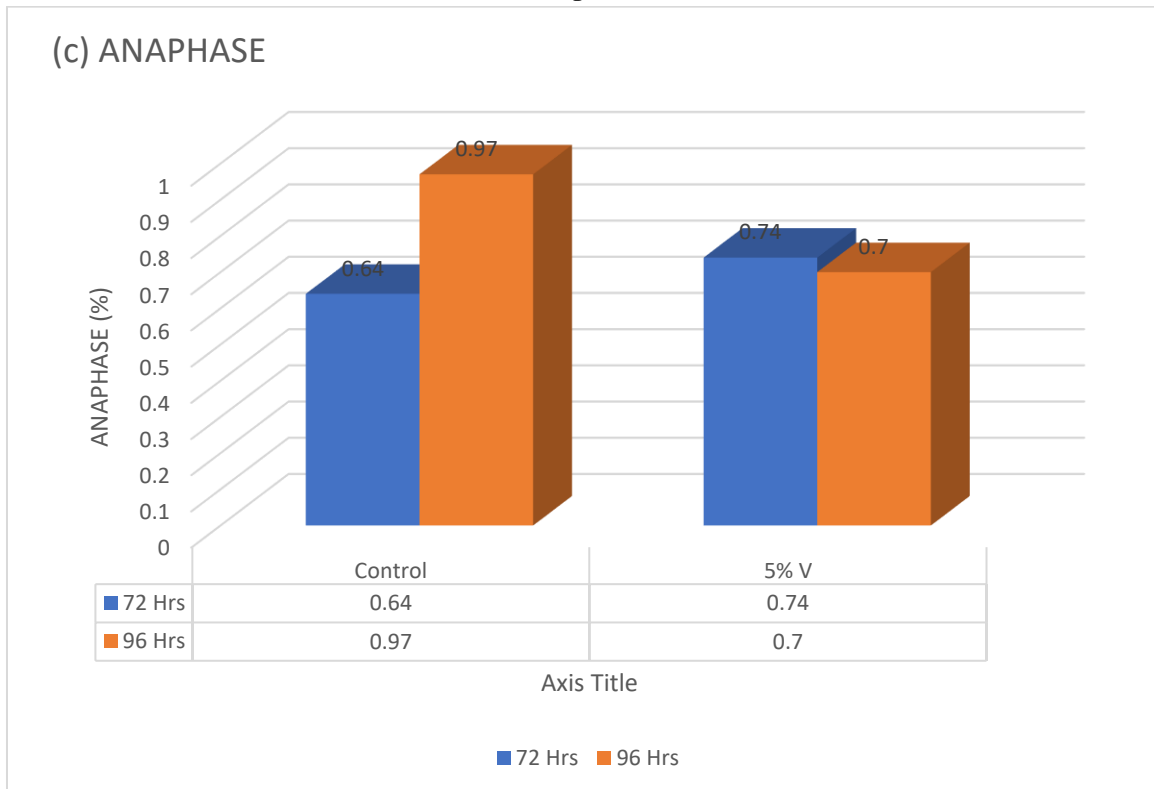


(B) PHASE DISTRIBUTION OF CONTROL AND VAN TULSI LEAF EXTRACT AT TWO DIFFERENT DURATION 72 HRS AND 96 HRS IN ONION ROOT TIP CELLS.





Anaphase



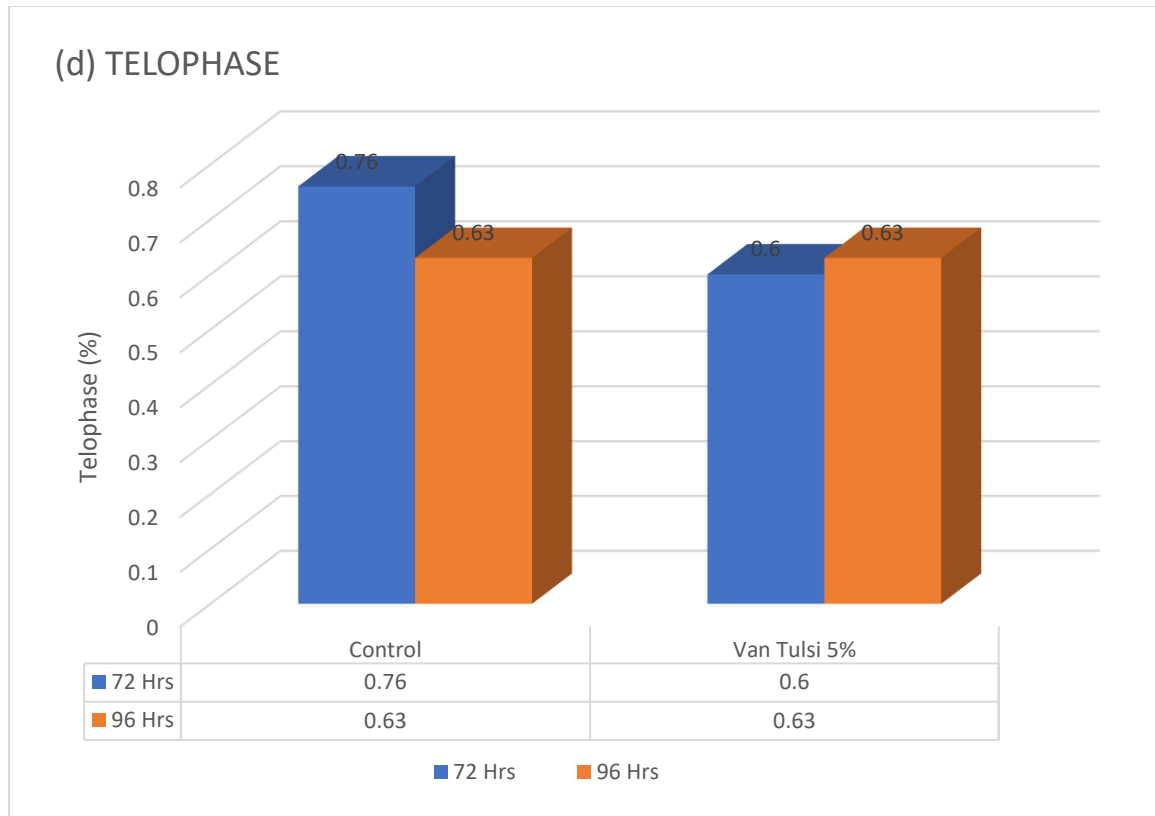


Figure 3: Histogram showing (a) Mitotic index (b) Phase distribution (Prophase; Metaphase; Anaphase; Telophase) in Onion Root Tip Cells at 72hrs and 96hrs in Control and Van Tulsi (5%) Leaf extract Treated group

RESULT AND DISCUSSION:

In 72hrs treatment duration there is no significant difference were found in mitotic index and phase distribution of Van Tulsi treated onion root tips cells than control group of onion root tips cells. This suggest that 72hrs concentration of Van Tulsi could not induce cytotoxicity.

In 96hrs treatment duration, the mitotic index significantly decreased from 34.63% to 22.00% and in phase distribution mitotic index of Prophase and Metaphase significantly decreased from 29.42 to 18.7% and 3.26 to 2.10%. This decrease in mitotic index was mainly due to a decrease in the population of cells belonging to Prophase and Metaphase (Table 1).

Similar results were observed [23]. Numerous studies have shown that wherever there is root growth inhibition in *Allium test*, there is also reduction in the number of dividing cells [24].

REFERENCE:

1. Pessoa L. M., Morais S. M., Bevilaque C. M., Luciano J. H., (2002). Antihelmintic activity of essential oil of *Ocimum gratissimum* Linn, and eugenol against *Heamonchus contortus*. *Veterinary Parasitolog*, 10:59–63.
2. Ramaiah M., Prathi A., Singam B., Tulluru g., Tummala l., (2019). A review on *Ocimum* species: *Ocimum Americanum* L., *Ocimum Basilicum* L., *Ocimum gratissimum* L. and *Ocimum tenuiflorum* L. *International Journal of Research in Ayurveda and Pharmacy*, 10 (3).
3. Bernell S. and Howard S. W., (2016). Use your words carefully: what is a chronic disease? *Front. Public Health*, 4:159.

4. Venuprasad M. P., Kandikattu H. K., Razack S., Khanum F., (2014). Phytochemical analysis of *Ocimum gratissimum* by LC-ESI – MS/MS and its antioxidant and anxiolytic effects South Afr. J. Bot., Le, 92:151–158.
5. Aguiyi J. C., Obi C. I., Gang S. S., Igweh A. C., (2000). Hypoglycaemic activity of *Ocimum gratissimum* in rats. *Fitoterapia*. 71:444–446.
6. Casanova L. M., da Silva D., Sola-Penna M., Camargo L. M., Celestrini D., Tinoco L. W., Costa S. S., (2014). Identification of chicoric acid as a hypoglycemic agent from *Ocimum gratissimum* leaf extract in a biomonitoring in vivo study. *Fitoterapia*. 93:132–141.
7. Ajayi A. M., Ben-Azu B., Onasanwo S. A., Adeoluwa O., Eduviere A., Ademowo O. G., (2019). Flavonoid-rich fraction of *Ocimum gratissimum* attenuates lipopolysaccharide-induced sickness behavior, inflammatory and oxidative stress in Mice. *Drug Res*. 69:151–158.
8. Umamageswari, A. and Kudagi B. L., (2016). Anti-inflammatory and analgesic properties of *Ocimum sanctum*: a comparative study using animal models. *International Journal of Basic & Clinical Pharmacology*, 4(5), 981–986. <https://doi.org/10.18203/2319-2003.ijbcp20150878>.
9. Offiah V. N., Chikwendu U. A., (1999). Antidiarrhoeal effects of *Ocimum gratissimum* leaf extract in experimental animals. *J. Ethnopharmacol*. 68:327–330.
10. Akara E. U., Emmanuel O., Ude V. C., Uche-Ikonne C., Eke G., Ugbogu E. A., (2021). *Ocimum gratissimum* leaf extract ameliorates phenylhydrazine-induced anaemia and toxicity in Wistar rats. *Drug Metab Pers Ther*. Apr 9;36(4):311–320. doi: 10.1515/dmpt-2020-0185. PMID: 34821128.
11. Shaw H. M., Wu. J. L. and Wang M. S., (2017). Antihypertensive effects of *Ocimum gratissimum* extract: angiotensin-converting enzyme inhibitor in vitro and in vivo investigation. *J. function. foods*. 35,68–73.
12. Melo R. S., Azevedo A. M. A., Pereira A. M. G., Rocha R. R., Cavalcante R. M. B., Matos M. N. C., Lopes P. H. R., Gomes G. A., Rodrigues T. H. S., Dos Santos H. S., Ponte I. L., Costa R. A., Brito G. S., Júnior F. E. A. C., Carneiro V. A., (2019). Chemical composition and antimicrobial effectiveness of *Ocimum gratissimum* L. Essential oil against multidrug-resistant isolates of *Staphylococcus aureus* and *Escherichia coli*. *Molecules*. 3864:2–17.
13. Nakamura C. V., Ueda-Nakamura T., Bando E., Melo Cortez A. F., Cortez D. A., Dias Filho B. P., (1999). Antibacterial Activity of *Ocimum gratissimum* L. Essential Oil, *Mem Inst Oswaldo Cruz* 94(5): 675-678.
14. Ugbogu O. C., Emmanuel O., Agi G. O., Ibe C., Ekweogu C. N., Ude V. C., Uche M. E., Nnanna R. O., Ugbogu E. A., (2021). A review on the traditional uses, phytochemistry, and pharmacological activities of clove basil (*Ocimum gratissimum* L.). *Heliyon*, Nov 25;7(11): e08404. doi: 10.1016/j.heliyon.2021. e08404. PMID: 34901489; PMCID: PMC8642617.
15. Mohr F. B., Lermen C., Gazim Z. C., Gonçalves J. E., Alberton O., (2017). Antifungal activity, yield, and composition of *Ocimum gratissimum* essential oil. *Genet Mol Res*.16;16(1). doi: 10.4238/gmr16019542. PMID: 28362991.
16. Chatterje A., Sukul N. C., Laskel S., Ghoshmajumadar S., (1982). Nematicides principal from two species of *Lamiaceae*. *J Nematol*, 14:118–20.
17. Chavan S. R. and Nikam S. T., (1982). Mosquito larvicidal activity of *Ocimum basilicum* Linn. *India J Med Res*, 75:220–2.

18. Faria T. D. J., Ferreira R. S., Yassumoto L., Roberto J., De Souza P., Ishikawa N. K., Barbosa A. D. M., (2006). Antifungal activity of essential Oil Isolated from *Ocimum gratissimum* L. (eugenol chemotype) against Phytopathogenic Fungi. Pp.867-871 ISSN 1516-8913, Londrina, PR-Brazil.
19. Joshi R. K., (2013). Chemical composition, in vitro antimicrobial and antioxidant activities of the essential oils of *Ocimum gratissimum*, Sanctum and their major constituents. Indian J. Pharmaceut. Sci. 75:457–462.
20. Mahapatra S. K., Roy S., (2014). Phytopharmacological approach of free radical scavenging and anti-oxidative potential of eugenol and *Ocimum gratissimum* Linn. *Asian Pac. J. Trop. Med.* 7S1:S391–397.
21. Bansal J; Kumar N; Rishabha M., Sharma P. K., (2013). Extraction and Evaluation of Tamarind Seed Polysaccharide as Pharmaceutical In situ Gel Forming System. *American-Eurasian Journal of Scientific Research.* 9. 1-5.
22. Akinboro A. and Bakeres A. A., (2013). Spermatotoxic, cytotoxic and genotoxic evaluation of aqueous extract of *Ocimum gratissimum* in albino mice. *Open Access Journal of Medical and Aromatic Plant* vol 4(1):10-14.
23. Kumari R., Kumari D., & Kumari A. R., (2019). Cytotoxic and genotoxic effect of profenofos on root tips cells of *Allium cepa* (Onion). *International Journal of Research Analytical Reviews* 6(2)676 – 680.
24. Rana P. K. and Kumai D., (2022) Cytotoxic effect of marigold (*Tagetes erecta*) leaf extract on root tip cells of onion (*Allium cepa*). *IJIRMF* 8(6).87-89.