

Synaptic Pruning in Schizophrenia: Manifestations and Antecedents

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Abstract:

Schizophrenia (SCZ) is a complex neurodevelopmental disorder characterised by structural and functional abnormalities in the brain. The synaptic pruning (SP) hypothesis, originally proposed by Irvin Feinberg (1982), suggests that excessive SP during adolescence contributes to the pathogenesis of SCZ. SP is a normal neurodevelopmental process wherein redundant synapses are eliminated to refine neural circuits. However, in SCZ, SP is aberrant and manifests as widespread grey matter depletion, reduced dendritic spine density, and cognitive deficits. This paper explores the structural and functional consequences of abnormal SP in SCZ and its potential antecedents, such as complement component 4 (C4) overexpression, neuroinflammation, NMDA receptor dysfunction, genes, and environmental deprivation and insults. Understanding the interplay between SP and schizophrenia pathophysiology may provide crucial insights into its neurodevelopmental origins and novel treatment strategies, especially for children showing early signs of psychosis.

Keywords: Cognitive, grey matter, schizophrenia, synaptic pruning

I. INTRODUCTION

Schizophrenia (SCZ) is a complex neurodevelopmental psychiatric disorder that affects how a person thinks, feels, and behaves. It is characterised by a range of symptoms broadly categorised as positive, negative, and cognitive. Positive symptoms refer to the presence of abnormal behaviours or experiences that are not typically seen in healthy individuals. These include hallucinations (auditory or visual perceptions without external stimuli); delusions (firmly held false beliefs despite contradictory evidence); disorganised thinking and speech (manifested as loose associations, tangentiality, and in severe cases, incoherence); and grossly disorganised or abnormal motor behaviour (ranging from childlike silliness to unpredictable agitation to catatonia). Conversely, negative symptoms involve a diminution or absence of typical behaviours and functions. These encompass avolition (lack of motivation), blunted affect (reduced emotional expression), asociality (social withdrawal), anhedonia (inability to experience pleasure), and alogia (poverty of speech). Cognitive symptoms of SCZ impact memory, attention, working memory, problem-solving, processing speed, and social cognition [1]. Not all symptoms need to be exhibited and the combination thereof varies among patients. Centuries of research on the aetiopathogenesis of SCZ have revealed it to be incredibly complex, with multiple intersecting hypotheses and models having been

proposed to explain it. One such hypothesis is the synaptic pruning hypothesis put forth by Irvin Feinberg [2].

During the early prenatal stages, the brain normally forms more synapses than it will require to function optimally. This overabundance allows the brain's circuits to be finely tuned to the demands of any environment it is exposed to. This fine-tuning is termed synaptic pruning (SP), a process responsible for the large-scale remodelling of the developing brain that culminates in the highly precise and efficient wiring that characterises mature neural circuits [3]. Neural activity, including that occurring in response to environmental stimuli, guides SP, such that less frequently used synapses are eliminated and more frequently used ones are preserved and strengthened. Such synaptic elimination (including in the prefrontal cortex [4]) happens primarily through phagocytosis by microglia, the resident macrophages of the central nervous system (CNS). Like cellular debris and pathogens, synapses that need to be removed are tagged (or opsonised) by molecules such as complement component 3. Microglia then recognise these tags and engulf the marked synapses.

The brain's neural circuitry is largely pruned during two developmental stages: the first two postnatal years and adolescence [5]. Abnormal pruning at both times is associated with several neuropsychiatric conditions [6–14]. More specifically, the SP hypothesis of SCZ states that excessive SP during adolescence contributes to the aetiopathogenesis of SCZ [2]. The hypothesis connects to abnormal SP several observations about SCZ patients, such as broad structural and functional abnormalities in their brains and the symptoms of psychosis typically emerging after the second wave of SP [2,15]. This paper explores the literature pertaining to aberrant SP in SCZ, including its structural and functional manifestations, and potential antecedents and mechanisms.

II. MANIFESTATIONS OF ABERRANT SYNAPTIC PRUNING IN SCHIZOPHRENIA

A. Smaller Brain Volume

Structural imaging research has indicated reduced whole brain volumes during the prodromal phase, at the onset of psychosis, and in chronic illness [16–20]. For instance, a meta-analysis involving over 18,000 subjects revealed that both medicated and antipsychotic-naïve patients exhibit decreased intracranial and total brain volumes, with the most pronounced reductions observed in grey matter (GM) structures such as the frontal and temporal lobes, cingulate and insular cortex, and the thalamus [21]. An eight-year longitudinal MRI study revealed that individuals with a high genetic risk for SCZ possess smaller whole brain, PFC, and temporal lobe volumes over time than those without genetic risk. Additionally, those who eventually developed SCZ exhibited even greater PFC volume reduction [22]. Smaller hippocampal volumes have also been observed in the early stages of the illness [23–24], consistent with the temporal lobe findings. Volume reduction in SCZ is particularly notable in cortical areas, whereas structural imaging studies of the basal ganglia [25–27] and cerebellum [28–30] have shown mixed results. GM depletion is considered the primary phenomenon underlying these brain volume reductions in SCZ patients [31–32].

B. Grey Matter Depletion

Over the course of brain maturation, regressive events such as SP reduce GM density; whereas concomitant progressive events such as myelination increase white matter (WM) density [33–34]. Excessively reduced GM has been observed widely across the cerebral cortex, especially in the frontal and temporal lobes of individuals at high risk for SCZ who eventually develop psychosis [22, 35–39], individuals experiencing their first psychosis episode [18, 40–43], and patients diagnosed with SCZ [18, 44–49]. Thompson et al. [50] conducted multiple MRI scans on 12 adolescents with SCZ and 12 without

over five years. They observed a gradual GM loss in those with SCZ, initially affecting the parietal association areas and later extending to the parietal, motor, temporal, and prefrontal regions.

Further, a longer duration of illness [51–52] and the severity of negative symptoms [44, 52] are correlated with lower GM volume in several areas including the left dorsomedial prefrontal cortex (PFC), right middle frontal cortex [51] frontotemporal region [44], the superior temporal gyrus, and anterior cingulate cortex [52]. Conversely, improved cognitive symptoms in domains such as working memory, attention, and psychomotor speed are observed in SCZ patients with a greater GM volume [51].

These abnormal GM reductions are attributed to exaggerated regressive brain maturational events such as SP because of the temporal [40] and spatial [42] coincidence of pathological and non-pathological cortical thinning in SCZ patients and controls, respectively. In other words, it is the frontal lobes, the areas pruned during adolescence and young adulthood, that display the most significant GM depletion in SCZ, whose symptoms are also known to emerge during adolescence and young adulthood. Furthermore, the overactivity of microglia, the cells that phagocytose synapses, also depletes GM [53–54].

While appropriate GM reductions are perturbed by synaptic over-pruning, the brain WM of SCZ patients, especially within the frontal and temporal lobes, also appears less dense and of poorer structural integrity [55–57]. Abnormal myelination has also been inferred, as measured by the grey-to-white matter contrast [58], with improvement correlating with the duration of antipsychotic intake [41]. Akin to SP, myelination also occurs in a posterior-to-anterior direction, with the frontal and temporal lobes myelinating during adolescence, among other life stages [59–60]. Once again, microglia seem to be involved in the aforementioned WM anomalies as activated microglia have been found near dystrophic and apoptotic oligodendrocytes, demyelinating and dysmyelinating axons in SCZ patients [61]. Oligodendrocytes are the glial cells mainly responsible for myelinating axons in the CNS. Other contributing factors include morphological abnormalities in oligodendrocytes and attenuated expression of oligodendrocyte- and myelin-related proteins in SCZ patients [61].

C. Dendritic Spine and Neuropil Depletion

Glausier & Lewis [62] ascribe smaller GM volumes in SCZ to neuropil shrinkage instead of neuronal death [63–65]. Neuropil is a region, most prominently occurring in the brain, that comprises unmyelinated axons, axon terminals, dendritic shafts, and dendritic spines. Particularly, dendritic spines are distinct morphological and biochemical structures that extend from the dendritic shafts of pyramidal cells in the cortex and hippocampus, among other brain areas. Spines are the substrate for 80–95% of excitatory synapses in the CNS [66–68], with each spine typically receiving one glutamatergic synapse [69]. Therefore, spine density marks the amount of excitatory input a neuron gets [70].

In SCZ, there is a notable reduction in spine density and size across various cortical regions, such as the striatum, the subiculum, the temporal, auditory, and prefrontal cortices [62, 71–78]. Notably, lower spine density is observed in layer 3 pyramidal neurons (PNs) of the dorsolateral PFC [73], an anomaly absent in deeper cortical layers, such as layers 5 and 6 [79]. Further evidence, although indirect, comes from experimental studies wherein reduced dendritic spines were linked to deficits in sociability, sensorimotor processing, working memory, and attention, also commonly seen in SCZ [80–81]. Such a decrease in dendritic spine density seems to result from a combination of overpruning, impaired spinogenesis, genes, and cytoarchitectural defects [82].

Dendritic spines have been implicated in the aetiogenesis of SCZ since they increase during childhood and prominently decrease during adolescence [83–87]. Moreover, dendritic spines have dynamic shapes and change form slightly throughout their existence [88]. Although the different shapes exist on a

continuum [89], two have been commonly studied in relation to SCZ and SP. Firstly, small, thin spines are transient and dynamic and, therefore, important for learning, which requires rapid plasticity via new connections [88, 90–91]. Secondly, large, mushroom spines, being more mature and persistent, participate in long-term activity-dependent synaptic plasticity and memory retention [92–93].

Studies on rats show that during adolescent SP, thinner, more immature spines and precursors thereof (filopodia) are the principal targets of elimination in the neocortex, sparing larger, more enduring spines [4, 94]. These findings align with the activity-dependent nature of SP that follows a use-it-or-lose-it principle [95–96]. Thus, it may be speculated that thin spines are selectively compromised in SCZ, but the reason for this is unclear. Indeed, two postmortem studies have found a preferential decrease in smaller spines, but these investigations are limited to the primary auditory cortex. One showed its association with altered calcium channel signalling involving the SCZ risk gene *CACNB4*, ultimately hindering spinogenesis [97]. Another associated it with reduced microtubule-associated protein 2 (MAP2) immunoreactivity, suggesting cytoskeletal abnormalities that may underlie synaptic dysfunction [98]. However, two more studies contradict these findings. Cai et al. [99] observed that a specific gene variant associated with SCZ risk preferentially decreased mushroom spine density in cultured primary hippocampal neurons of rats. Sánchez-González et al. [100] found increased thin spine density in frontal cortex PNs in a rat strain presenting schizophrenia-like traits, reflecting immature synaptic states.

Additionally, Gonzalez-Burgos et al. [101] obtained findings that contrast with the theory that smaller spines are preferentially eliminated during adolescent SP. They showed that in layer 3 PNs in monkeys' DLPFC, weaker synapses (with small spines) get eliminated just as much as stronger synapses, thus supporting uniform pruning across the entire distribution of synaptic strengths. Given these findings, they inferred that the selective elimination of small spines, as observed by MacDonald et al. [97] and McKinney et al. [98], suggests an abnormal pruning process where strong synapses are eliminated normally while weak synapses are eliminated excessively, which disrupts the normal distribution of synaptic strengths and reduces the excitatory drive to layer 3 PNs. Thus, there is an apparent lack of consensus concerning the morphological type of dendritic spines in SCZ and its relation with SP.

D. Cognitive Symptoms

Cognitive symptoms, including deficits in attention, psychomotor speed, information processing, learning, memory, abstract thinking, planning, and problem-solving, are a widespread and disruptive domain of SCZ symptomatology [102]. These denote a trait marker for SCZ since they appear even before the onset of the illness [103–104]. They seem to stem from a combination of disrupted neuroplasticity (NP) [105–112] and abnormal brain connectivity [114–115]. This is reflected in functional neuroimaging findings in SCZ such as aberrant frontotemporal activity and connectivity, thought to contribute to deficits in verbal memory and encoding; abnormal activation in sensory pathways in response to complex stimuli, indicating faulty sensory integration; and unbalanced top-down and bottom-up processes during information-processing (summarised in Boland & Verduin [1]). Conversely, interventions such as cognitive remediation therapy (CRT) target NP and neural rewiring, improving cognitive performance, both globally and in specific domains, such as social cognition, verbal working memory, and daily functioning [42, 116–119]. These positive outcomes are also reflected in neuroimaging studies reporting increased frontal and temporal activation, enhanced resting-state functional connectivity, and lesser GM loss post-intervention, accompanied by improved performance on various cognitive tasks [118, 120].

Synaptic Pruning and Neuroplasticity. We present four lines of evidence that relate to both NP and SP. Firstly, long-term potentiation (LTP) and long-term depression (LTD), at two complementary scales,

underlie NP and SP. LTP strengthens synaptic connections to enhance inter-neuronal communication by promoting spine turnover and synapse stabilisation [121- 122]. On the other hand, LTD weakens synaptic connections by inducing spine shrinkage and loss, increased varicosities turnover, and increased separation of varicosities from dendritic spines [121, 123–125]. At an immediate and microcircuit level, LTP and LTD allow NP by adjusting synaptic strength at individual synapses, enabling rapid, activity-dependent changes critical for processes like learning and memory [126–127]. At a developmental and microcircuit level, these phenomena play a vital role in SP and shaping neural circuitry [128].

The few investigations on LTP in SCZ have reported impaired LTP-like plasticity in the patients' motor [129–131] and visual [132] cortices in response to non-invasive stimulation. Two studies particularly found impaired focal LTP-like plasticity rather than the non-focal type, indicating subdued plasticity and signal integration at the synapse level [129–130, 133]. Further, one study established a relationship between subdued LTP-like plasticity in the motor cortex and patients' deficits in rotary pursuit motor learning [131]. Such faulty LTP is seen as a result of dysfunction of glutamatergic and dopaminergic neurotransmission, N-methyl-D-aspartate (NMDA) receptors, and inhibitory regulation comprising the gamma-aminobutyric acid (GABA)-synthesising enzyme glutamic acid decarboxylase and GABAergic neurotransmission [111, 129, 134–138]. Abnormal density, trafficking, and subunit expression of α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors may also play a role, although the findings are inconclusive [139]. Even fewer investigations on LTD in SCZ also indicate abolished LTD-like plasticity [130, 140].

The second line of evidence pertains to dendritic spines, which contain NMDA and AMPA receptors, making them the anatomical substrate for NP. As explained in the previous subsection, these appear to be depleted in patients with SCZ, likely due to excessive elimination or pruning. Thirdly—and more generally—the 108 SCZ-associated genetic loci analysed by Ripke et al. [113] are those that encode synaptic proteins essential to synaptic plasticity, such as NMDA, AMPA, the voltage-dependent calcium channel, and dopamine receptor D2. Lastly, environmental factors also influence synaptic plasticity and SP: early-life stress and cognitive deprivation form the two common themes among the environmental risk factors of SCZ, as explained later. In both animal and human studies, these are linked to quicker brain maturation and reduced NP, further tied to cognitive impairments akin to those seen in SCZ [141].

Synaptic Pruning and Neural Wiring. Enfeebled LTP and LTD would mean that the required synaptic connections are not formed/strengthened, and the trivial ones are not removed/weakened. This may obfuscate the utility of specific brain networks, a crucial determinant for the process of SP. Normally, SP optimises circuitry, manifesting as superior functional outcomes and signal-to-noise ratio (SNR) of neural transmission [142–145]. However, in the brains of SCZ patients, such disturbed SP may explain the widespread hypoconnectivity observed within and among several key networks using resting-state functional connectivity, namely the default mode network, auditory network, core network, somatomotor network, salience network, and the self-referential network [114]. These findings support the dysconnectivity hypothesis of SCZ by Stephan et al. [146].

This explains the abnormal functional neuroimaging findings obtained in SCZ patients, such as faulty sensory integration, and top-down and bottom-up processing during information processing [1]. Specifically, the following have been reported: abnormal activation in the dorsal visual pathway in response to complex stimuli conditions, despite normal activation in basic sensory and motor regions in response to simple stimuli; faulty inhibitory P300 for a no-go condition, indicating reduced inhibition by the PFC; in event-related potential (ERP) studies with oddball tasks, aberrant activation in patients

correlates with symptom severity: attenuated response to targets is thought to reflect insufficient top-down activation of target circuitry, whereas attenuated evoked response to novel distractors may reflect overprocessing of bottom-up events [1]. Additionally, a lower SNR has been observed, with dopamine, glutamate, and GABA dysfunction as possible antecedents [147], which are already known to be involved in the aetiology of SCZ. This reduced SNR manifests (in SCZ patients and, to a lesser extent, in their non-diagnosed siblings) as insufficient prefrontal activation during cognitive tasks in fMRI data and variability in prefrontal ERPs in EEG data [147]. Further, [148] proposed neural network abnormalities in SCZ due to pathological SP as the reason for a conflict between the excitatory and inhibitory processes in the brains of SCZ patients. This was manifested in their EEG results as either a simultaneous increase or decrease in ERP latency and amplitude when shown neutral and threatening images.

III. POTENTIAL ANTECEDENTS TO ABERRANT SYNAPTIC PRUNING IN SCHIZOPHRENIA

A. Immune and Microglial Dysfunction

Immune Dysregulation and Neuroinflammation Drive Microglial Overactivation: Since microglia are the resident macrophages of the CNS, their role in synaptic elimination also constitutes the neuroinflammation hypothesis of SCZ, which posits that dysregulation of the immune system and inflammatory processes play a significant role in the pathophysiology of SCZ [149]. SCZ risk is elevated by prenatal and childhood infections [150], and autoimmune disorders independently and in combination with infections [151]. Next, inflammatory mediators are involved in SP [152] and are associated with SCZ [148, 153–156]. Thus, the link between neuroinflammation and SCZ might be mediated by microglial overactivation and the resulting hyperfunctioning, including aggressive SP [157], indicated by activated microglia found in postmortem studies of SCZ patients [158–160].

C4 Overexpression Aggravates Microglia-Driven Synaptic Elimination: Genome-wide association studies (GWAS) have uncovered common genetic variants associated with a heightened risk of SCZ [161]. The most robust findings involve the major histocompatibility complex (MHC) locus on chromosome 6 [113, 162]. Particularly, repeated copy numbers of the complement component 4 (C4) gene result in increased expression of the C4A (but not C4B) isoform of C4, an immune system signalling molecule [162, 163]. C4A participates in SP by activating complement component (C3), which opsonises dendritic spines of weak or less active synapses for phagocytosis by microglia [164]. In vitro models have shown that microglia-mediated SP and synaptic engulfment are more aggressive in SCZ when either the microglia or neurons in cell culture are derived from patients, in comparison to healthy controls [165]. This aligns with observations of reduced spine density in SCZ, suggesting a mechanism of excessive synaptic elimination.

Moreover, C4A expression in humanised mouse models [166] and patient-derived cultures [87, 167] directly correlates with increased microglial engulfment and neuronal complement deposition. However, although C4A concentration is elevated in the cerebrospinal fluid (CSF) of first-episode psychosis patients who develop SCZ, this increase is not fully explained by genetic risk variance at the C4 locus [168]. Contributing towards a more comprehensive explanation, Gracias et al. [168] discovered that cytokines interleukin (IL)-1 β and IL-6 in patient-derived cellular models and IL-1 β in patient-derived CSF selectively enhance C4A mRNA expression in neurons, even after controlling for genetically predicted C4A mRNA expression. Prasad et al. [169] also found a positive correlation between IL-6 in the peripheral blood and catabolites of membrane phospholipids in the thalamus of the brains of early-course SCZ

patients. These catabolites were thought to reflect neuropil depletion, likely due to excessive pruning. They further cite evidence indicating that smaller thalamic volume is associated with negative and positive symptoms and hindered cognitive performance. Additional indirect evidence (summarised in Pearson & Iadecola [170]) comes from studies that have established associations between maternal infections, interleukins, and SCZ.

Potential Treatments: Studies on potential treatment drugs consistent with these mechanisms have shown that minocycline, an antibiotic, reduces microglial engulfment of synapses and thereby reduces the risk of SCZ [89] as well as negative and cognitive symptoms [171]. Previous meta-analyses had also demonstrated the drug's effectiveness and safety in treating the negative symptoms ([172–173]; cf. Deakin et al. [174]; see also Inta et al. [175]). Wangemann [166] also found that oridonin, a targeted NLRP3 inflammasome inhibitor, exhibits anti-inflammatory properties in human induced pluripotent stem cells (iPSC)-derived microglia. It suppressed synaptosome phagocytosis and enhanced synaptic density in C4A overexpressing iPSC-derived cortical neurons when co-cultured with microglia. Suppression of microglial activity through anti-inflammatory agents also enhances the antipsychotic effect for treating negative and cognitive symptoms [171, 176–178]. Furthermore, various antipsychotic agents attenuate microglial secretion of pro-inflammatory substances such as nitric oxide and interleukins [179–183].

B. Excitatory-Inhibitory Dysfunction

Glutamate Surge in the Cortex Exacerbates Synaptic Pruning: Glutamate and GABA are the brain's primary excitatory and inhibitory neurotransmitters, respectively. The glutamate hypothesis of SCZ posits that hypofunction of the NMDA glutamate receptor (NMDAR) causes hypoactivation of GABAergic interneurons that express parvalbumin (PV+ interneurons), consequently producing disinhibition of excitatory glutamatergic PNs, which contributes to SCZ symptoms [184–186]. PV+ interneurons are fast-spiking and synchronise the activity of PNs, which constitute about 80% of all cerebrocortical neurons and generate the gamma band oscillations necessary for optimal cognitive function. Parellada & Gasso [82] review several lines of evidence that align with the glutamate hypothesis (see also Howes & Shatalina [187]), including the psychotomimesis induced in healthy subjects by NMDAR antagonists such as ketamine and phencyclidine and dysfunction of the PV+ interneurons in the PFC and hippocampus. Additionally, Orhan et al. [188] reported elevated kynurenic acid, an endogenous NMDAR antagonist, in patients' brains, which decreased neuronal activity and promoted microglial synaptic elimination.

Parellada & Gasso [82] also combine the glutamate hypothesis with the well-known dopamine hypothesis of SCZ by proposing that aberrant excitation of PV+ GABAergic inhibitory interneurons triggers (a) a disinhibited and excessive glutamate release by glutamatergic cerebrocortical PNs; and (b) hyperdopaminergic subcortical areas, for which the disinhibition of long-range pyramidal cortical projection to the ventro tegmental area may also be responsible. They further explain that the glutamate surge and calcium overload through glutamate receptors can initiate excitotoxicity and rapid dendritic pruning (or non-lethal synaptic apoptosis) by locally activating the mitochondrial apoptosis pathway and the caspase-3 cascade, resulting in a reduced density of dendritic spines. Parellada & Gasso [82] cite evidence showing that apoptotic dendrites are more likely to be eliminated by microglia because they release certain “find me” (e.g., glutamate levels) and “eat me” (e.g., complement cascade proteins such as C1q and C3) signals for the microglia. Thus, the hyperglutamatergic state and its consequences are tied to reduced NP, dysfunctional brain regions, and the negative and cognitive symptoms of SCZ.

Since dendritic spines form the principal microstructure of glutamate synapses [189] and their density is often reduced in the brains of SCZ patients [73], this could initiate the vicious cycle of NMDAR—and

therefore GABAergic interneuron—hypoactivation, followed by the uninhibited glutamate surge in the cortex, and excessive spine elimination or death. The possible antecedents to NMDAR dysfunction in SCZ are outside the scope of this paper, but research indicates a genetic basis [190, 113] in addition to a controversial auto-immunological basis [191–192].

Besides modulating the activity of GABAergic interneurons, a general function of NMDAR activation is to control cytoskeletal dynamics and the expression of structural proteins and neurotrophins like brain-derived neurotrophic factor (BDNF) that aid in synapse stabilisation, and to recruit AMPA receptors to help maintain spine integrity [193–194]. Therefore, alterations to these receptors may compromise the structural integrity of dendritic spines and reduce their activity levels, making them more susceptible to either death or elimination by microglia.

Disrupted GABA Neurotransmission May Drive Synaptic Pruning: Along with NMDAR, the GABA receptors form a crucial part of the excitatory-inhibitory balance in the brain. The role of GABA_A receptors—particularly the $\alpha 4\beta\delta$ isoform—in SP was demonstrated by Smith et al. [195], who injected its orthosteric agonist, gaboxadol, into male mice for 14 days, starting at the onset of puberty. They observed a 50% reduction in the dendritic spine density in layer 3 pyramidal cells of the prelimbic PFC and impaired temporal order recognition—a PFC-dependent cognitive deficit reported in SCZ. The $\alpha 4\beta\delta$ isoform is found on extrasynaptic sites in the forebrain and partakes in tonic inhibition, wherein the receptor isoform, when activated, remains open for a longer time, providing a lasting inhibitory tone [196]. Tonic inhibition is crucial for synaptic plasticity, neurogenesis [197–198], and cognitive functions [199–200]. The reduced spine density observed by Smith et al. [195] may be attributed to the increased probability of elimination of inhibited and relatively inactive neurons [201–203]. This may also be why administering NMDAR antagonist MK801 [204] and genetically deleting the NR1 subunit of NMDARs [205] reduces spine density in cortical PNs, since activation of NMDARs on a neuron excites the neuron. Furthermore, similar to how gaboxadol elicited impaired temporal order recognition, NMDAR antagonists also elicit the positive, negative, and cognitive symptoms of SCZ [206–207]. However, whether gaboxadol, like NMDAR antagonists, selectively disrupts GABA-related functioning [208–209] or also directly impacts glutamate neurotransmission is unknown. Moreover, the cortical PNs affected by NMDAR antagonists and gaboxadol also extend connections to the ventral tegmental area—a crucial part of the mesolimbic dopamine pathway that is thought to produce excessive dopamine and positive symptoms in SCZ.

Rabinovitch et al. [210] put forth a model suggesting that the genetic predisposition to SCZ leads to an overproduction of GABAergic neurons, possibly prenatally. However, in the adult brain, either (a) the excitatory-inhibitory (E/I) balance is skewed towards glutamatergic neurons due to the over-pruning of GABAergic neurons by homeostasis-maintaining glial cells; or (b) the E/I ratio is balanced but the power of both neuron types is diminished, a result of the glial cells also over-pruning glutamatergic synapses to re-establish the equilibrium disrupted by E/I imbalance towards glutamatergic power. The authors also propose a mathematical model and graph to represent this process.

Potential Treatments: Given that the hypoactivation of NMDAR contributes to SCZ symptoms, glutamatergic drugs or NMDAR agonists have been proposed as treatments. Orhan et al. [188] found that the decrease in spine density caused by kynurenic acid in excitatory neurons developed from iPSCs was reversed by adding NMDA and the endogenous co-agonist D-serine. A Cochrane review also reported a moderate effect of D-serine and glycine when complementing antipsychotics but pointed out the inconsistent data and small sample size [211]. However, D-cycloserine, a partial agonist of NMDAR's glycine site, did not improve symptoms. Reduced levels of D-aspartate, an endogenous NMDAR agonist

[212]; quinolinic acid, an endogenous NMDAR agonist [213]; and LY379268, an agonist of mGluR2/3, a group II metabotropic glutamate receptor [214], have also been implicated in SCZ. Engel et al. [214], in mice models, showed that LY379268 restores the excitatory-inhibitory balance disrupted in SCZ and that it does so with efficiency comparable to olanzapine. Similarly, [215] genetically and pharmacologically elevated D-aspartate levels in mice. This increased NMDAR activity, improved cognitive and memory functions, and attenuated schizophrenia-related deficits in sensorimotor gating induced by amphetamine and MK-801. In light of studies implicating deficient GABA neurotransmission in SCZ and its upregulation in the regulation of the onset and duration of pre-adolescent SP, preliminary evidence suggests that tiagabine, a GABA reuptake inhibitor that blocks the GABA transporter 1 (GAT-1), may improve cognitive symptoms in early course of SCZ by modulating PFC functions, as shown in fMRI scans taken as subjects performed working memory tasks [216–217]. Although the aforementioned drugs by themselves may not be enough to treat a disorder as multifaceted as SCZ, further translational and clinical research is nevertheless needed to determine their therapeutic effectiveness.

C. Genes

RELN. Arkhipov et al. [148] implicate the RELN gene in synaptic elimination. The RELN gene codes for reelin, an extracellular matrix protein that modulates the formation of neural systems and synaptogenesis through its trophic and signalling role [218–219]. In addition, reelin plays an active role in regulating glutamatergic neurotransmission and aids in the development of dendritic spines, which helps maintain optimal synaptic plasticity [218, 220–221]. The binding of reelin to receptors triggers cascades contributing to dendritic proliferation, dendritic growth, and branching in the cortex [222–226]. Arkhipov et al. observed abnormally high levels of reelin in patients' peripheral blood due to decreased methylation (repression) of the promoter region of the RELN gene. Given that methylation in peripheral blood is inversely proportional to that in the brain [227], they state that reelin levels must be reduced in patients' brains (previously confirmed by Impagnatiello et al. [228]) due to the hypermethylation of the RELN gene. They theorise that the reduced reelin may result in indirect connections, insufficient synaptic elimination, and abnormal neural processing evident in their EEG data, which depicted an imbalance of excitation and inhibition processes in the brain. However, part of the results obtained by Arkhipov et al. [148] are inconsistent with previous studies that found a reduction in RELN mRNA expression [229] and heightened methylation of the RELN promoter [230] in SCZ patients' peripheral blood (see also Markiewicz et al. [231])

Boksa [232] purports that the reduction in dendritic spine density in reelin-deficient rodents may result from reelin's role in cytoskeleton stability. She explains that “phosphorylation of the F-actin binding protein, n-cofilin, inhibits its actin-severing activity, leading to stabilisation of the cytoskeleton, & it has been shown that signalling by reelin is required for n-cofilin phosphorylation [233]. Thus, in patients, reduced reelin may reduce cytoskeleton stability & dendritic spine density via a decrease in n-cofilin phosphorylation” (p. 2). Unsurprisingly, decreased reelin expression in the brain has been considered a vulnerability factor within the 2-hit model of SCZ aetiology [228]. Moreover, RELN supplementation may have a trophic effect on synaptic plasticity as it helps recover deficits in dendritic spine density, synaptic function, and cognitive performance that are associated with reduced RELN expression [234].

RhoGTPases. Certain genes & their respective proteins that have been implicated in SCZ are also involved in dendritic spine stability and elimination. For example, a family of small RhoGTPases (RhoA, Rac1, Cdc42) regulates the actin cytoskeleton that determines the morphology and, therefore, the synaptic function of dendritic spines. The mRNAs for Cdc42 and Duo (orthologous to Kailrin-7) are reduced in the

DLPFC of SCZ patients, and their expression correlates with spine density in the same region [235]. In the same study [235], the mRNA expression of Cdc42 and Duo was unaltered in monkeys chronically exposed to the antipsychotics haloperidol or olanzapine.

DISC1. Hayashi-Takagi et al. [236] explain the interactions among various proteins that influence dendritic spine stability, and ultimately, the probability of their elimination. Kalirin-7 (Kal-7) binds to and activates Rac-1, which is needed for spine growth, but its overexpression (and underexpression; see [235]) hampers spines. DISC1 (Disrupted-in-Schizophrenia 1), often associated with SCZ, prevents Kal-7 from binding to Rac-1 by confining the former's access to the latter, while the expression of DISC1 itself is inhibited by NMDAR activation. DISC1 is required for appropriate spine maintenance, but its overexpression leads to spine shrinkage. Thus, both over- and under-expression of DISC1 can be implicated in SCZ. Such interaction of DISC1 with proteins governing the actin cytoskeleton may account for the loss of dendritic length & spine density seen in DISC1 mutant mice [237]. Thus, the gene affects NP [238–239], which underlies the cognitive symptoms of SCZ, as explained previously. Various polymorphisms of DISC1 have been associated with SCZ [240–243]. DISC1 acts as a scaffold protein, and (a) as shown by Hayashi-Takagi [236], regulates many other proteins governing signalling and cytoskeletal structure; (b) regulates neurodevelopmental processes such as neurogenesis, migration, neurite outgrowth, and spine development; and (c) is involved in dopamine neurotransmission, which is generally aberrant in SCZ [244]. Since Bellon et al. [245] found that monocyte-derived-neuronal-like cells (MDNCs) from medicated patients prune more primary neurites after exposure to dopamine, DISC1 may play an indirect role in SP in SCZ. Besides DISC1, several other genes associated with SCZ are involved in overlapping processes such as SP, synaptic plasticity, and dendritic spine dynamics. These include Neuregulin 1 (NRG 1), Akt1, CACNA1C, and DTNBP1 [106].

DLG1, NOS1, THBS4, FADS1. Cocchi et al. [246] identified proteins associated with pruning by reviewing literature from 1995 to 2013 and using databases like PubMed and GeneMania. Genetic association with SCZ was tested using a sample of 9,490 subjects from the Psychiatric Genomics Consortium. Their genetic analyses found the following four genes to be relevant in SP and SCZ: DLG1 (Discs, large homolog) subserves synaptogenesis, signal transduction, cell proliferation, adherens junction assembly, and alteration of neuronal environment through glutamatergic and astrocyte activation. NOS1 (Nitric oxide synthase) codes for an enzyme that produces nitric oxide, which acts as a neurotransmitter and regulates neuron environment modifications and pruning. The gene is also associated with structural changes in the brain in SCZ. THBS4 (Thrombospondin) encodes an extracellular adhesive glycoprotein involved in cellular interactions, including migration (which is involved in pruning), neuronal development, and response to CNS injury. The protein also binds to ECM components important in SCZ and pruning. FADS1 (Fatty acid desaturase) catalyses the biosynthesis of highly unsaturated fatty acids, maintaining membrane structure and playing a role in pruning and SCZ. Through cell line and regional brain expression analysis, these genes were found to be primarily expressed in (a) the neuropil, to which the brain volume shrinkage found in SCZ is usually attributed [65, 261–263]; and (b) hippocampus, where notable differences manifest early in life between normal individuals, those at-risk, and those who will be diagnosed with SCZ in the future [264–265].

MHC. Certain other genes have also been recently linked to SCZ because of their role in microglial SP. The MHC on chromosome 6 forms the genetic platform for innate and adaptive immune processes. Its relation with SCZ has been explained in a previous section. The MHC encompasses the prominent human leucocyte antigen (HLA) region, whose variations are associated with SCZ [247–250]. Such variations

are also linked to a smaller thalamus [251], which further correlates with negative and positive symptoms [252] and cognitive performance [253]. Marco et al. [8] observed that the HLA-C and HLA-DRA genes showed significantly more single-nucleotide polymorphisms in 24% and 17% of the sample of SCZ patients, respectively. These genes involved in immunity and inflammation can alter microglial functions, potentially affecting neuronal and axonal remodelling, apoptosis, and SP. This harkens back to the neuroinflammation/immunity theory of SCZ described before.

Kynurenic Acid. Also mentioned before, Orhan et al. [188] found that kynurenic acid, an endogenous NMDAR antagonist usually elevated in the brains of SCZ patients, decreases neural activity and increases microglial SP. Additionally, by integrating large transcriptomic and genetic datasets, they demonstrated that the gene co-expression networks for KYNA-producing enzymes, namely the kynurenine aminotransferases (KATs), are independently enriched for genes involved in synaptic activity (including those coding for NMDAR subunits and $\alpha 7nAchR$) and common genetic risk variants for SCZ.

CYFIP1. The CYFIP1 gene, located in the 15q11.2 neurodevelopmental risk locus, encodes a protein that regulates actin cytoskeletal dynamics and is involved in dendritic complexity maintenance and mature spine stabilisation; it is also associated with SCZ [254]. Sheridan et al. [165] investigated its role in human microglial function by generating multiple CRISPR knockouts in patient-derived microglia models. They found that loss of CYFIP1 significantly reduced synaptic vesicle phagocytosis, altered microglial morphology to a more ramified (resting or surveillant) state, and decreased microglial motility. Drew et al. [255] also reported that conditional knockout of CYFIP1 in mice microglia led to impaired surveillance due to reduced mobility. However, they also reported decreased process ramification and increased activation marked by elevated CD68 expression. Additionally, CYFIP1 deletion or haploinsufficiency in mice results in defective neuronal morphology, plasticity and connectivity [254, 256–260].

D. Environmental Factors

Psychosocial. Among the most well-known postnatal environmental risk factors for SCZ are psychosocial. These include low socioeconomic status, first- or second-generation migrant status, social and material deprivation, social adversity, stressful life events, and adverse childhood experiences such as emotional, physical, and sexual abuse, mental illness in household members, bullying and collective violence, and emotional neglect [2, 15, 266]. Sheridan & McLaughlin [267] conceptualise such adverse experiences on two dimensions: threat and deprivation, which we shall consider as the primary themes among these overlapping factors. Although there is a paucity of studies that directly correlate these risk factors with markers of SP, specifically in SCZ patients, the impact of threat and deprivation on neurodevelopment—particularly SP—has been widely studied, allowing only indirect inference of associations between the aforementioned risk factors and SP.

Deprivation is defined by Sheridan & McLaughlin [267] as “the absence of expected environmental inputs in cognitive (e.g., language) and social domains as well as the absence of species- and age-typical complexity in environmental stimulation” (p. 13). Such deprivation may be experienced by children, preteens, and adolescents exposed to limited educational and recreational resources due to poverty; and few or poor social interactions due to discrimination, language barriers, or parental/caregiver neglect. Given the importance of neural activity for activity-dependent SP [3], both animal and human studies have shown that such deprivation accelerates and intensifies synaptic elimination, resulting in markedly reduced synapses per neuron, dendritic branching, and cortical thickness [141, 268–270]. As discussed before, these structural changes co-occur with deficits in learning and memory also found in SCZ patients.

A threat, according to Sheridan & McLaughlin [267] involves “the presence of an atypical (i.e., unexpected) experience characterised by actual or threatened death, injury, sexual violation, or other harm to one’s physical integrity” (p. 13). SCZ risk factors such as social adversity, community violence, stressful life events, and physical or sexual abuse, combined with the resulting stress response, may confer significant deviations from normal neurodevelopment, which align with results from neuroimaging data from SCZ patients. Exposure to threat, stress, and fear has been linked, in both animal and human models, to faster brain maturation [2, 141, 267]; reduced hippocampal volume, dendritic branching, and spine density in the hippocampus and PFC [141, 267]; blunted LTP in the hippocampus [267]; and aggravated microglial synaptic elimination in the PFC (but attenuated in the hippocampus [271–272]) through microglial overactivity and elevated C3 [274]. Exposure to early-life adversity and deprivation also impairs cognitive flexibility and associative learning [268, 270, 273], deficits that are also observed in SCZ patients [275–276] and may be a consequence of heightened SP at the cost of synaptic plasticity. Early childhood adversity can also make individuals more sensitive to stress during adolescence (see Smith & Pollak [277] for a detailed discussion), amplifying its deleterious effects on SP and contributing to the onset of SCZ. Furthermore, besides the valence of these psychosocial experiences, their frequency also influences SP. Chronic and repeated exposure to the same set of stimuli also trades synaptic plasticity with SP as the brain’s circuits refine to handle the (unvarying) demands of the environment [141]. During early development, inhibitory neurotransmission progressively increases [278]. The heightened plasticity characteristic of a critical neurodevelopmental period concludes when PV+ inhibitory interneurons mature and perineuronal nets (PNNs) form [279]. Exposure to early-life adversity decreases the number of PV+ interneurons and later leads to increased PNN accumulation around these cells, indicating reduced plasticity [280–281]. A reduction of PV+ interneurons was also found in the PFC of SCZ patients in a meta-analysis of post-mortem studies [282]. Moreover, as discussed earlier, Parellada and Gassó [82] theorised that the hypoactivation of said interneurons plays a crucial role in heightened synaptic elimination and the aetiology of SCZ.

Immunological. Another class of SCZ risk factors implicates the immune system, encompassing prenatal infections and prenatal exposure to maternal infections, the risk for which may be elevated by urban upbringing and gestation in the winter, also considered SCZ risk factors [283]. This association may be explained by immune system activation (including induction of immunity-related genes such as the MHC; [284]), resulting in heightened microglial reactivity and, subsequently, microglial engulfment of synapses [285–288]. In addition, increased pro-inflammatory genes in the cortex and hippocampi of SCZ patients may predispose their microglia to abnormally high activation [285]. A more permeable blood-brain barrier, microglial activation, and increased IL-6 and IL-1 β (also implicated in SP, as mentioned previously) may mediate the relationship between maternal immune activation and SCZ [170]. IL-1 β activates the pro-inflammatory M1 state of microglia, also triggered following neuronal injury ([289]; thereby also partly explaining the relationship between foetal hypoxia or other obstetric complications and SCZ risk [283, 290–291]). The dominance of the M1 state, coupled with an extended inflammatory response, results in the overproduction of pro-inflammatory cytokines and reactive oxygen species, ultimately causing synaptic loss and neuronal death [292]. Further, consistent with the diathesis-stress hypothesis of SCZ, perinatal activation of microglia may also make them vulnerable to overactivation by stressors experienced later in life, ultimately encouraging a shift to a pro-inflammatory M1 state [289].

In addition to disease and injury, social stress can also trigger the immune system and microglia-mediated SP in frontocortical and limbic circuits [293–295], whereupon the psychosocial and immunological risk

factors converge. Social stress activates the hypothalamic-pituitary-adrenal (HPA) axis and the sympathetic nervous system, both of which influence microglia and their reaction to stress [296–297], including heightened reactivity to later stress due to prior stress [298]. The adrenal cortex releases glucocorticoids in response to HPA axis activation. Since microglia express glucocorticoid receptors, high glucocorticoid levels can activate microglia, while mediators released by activated microglia can stimulate the HPA axis, producing a vicious cycle [299]. Further, elevated glucocorticoid levels correlate with the release of pro-inflammatory cytokines such as the interleukins [298], whose role in SP and SCZ was explained previously. At least two mechanisms link stress, microglia, and SP [298]. First, CX3CR1, the fractalkine receptor on microglia, is increasingly expressed in the cortex as a result of stress, while CX3CR1 knockout in mice enhances microglial engulfment and prevents further stress-induced modulation. Its deletion disrupts hippocampus-mPFC connectivity, impairs social interaction, and links microglia-driven SP to neuropsychiatric disorders. CX3CR1 is significantly downregulated in SCZ patients' blood and brain [300]. Second, and as stated previously, C3–C3R signalling tags synapses for microglial engulfment. Chronic stress elevates C3 and C3R levels, accelerating synaptic elimination [298]. Additional neurobiological effects of chronic stress include loss of dendritic spines and synapses, and these effects are associated with stress-induced cognitive-behavioural impairments such as anhedonia and working memory deficits (summarised in [156]).

Treatments. Several interventions for SCZ, such as cognitive remediation therapy (CRT), cognitive enrichment therapy (CET), and social skills training, aim to alleviate cognitive deprivation and promote synaptic plasticity. In early-course psychosis patients, CET preserves GM volume in the left hippocampus, parahippocampal gyrus, and fusiform gyrus and these neuroprotective effects correlate with improved cognitive outcomes [301]. CRT improves verbal fluency and increases hippocampal volume [302]. It is also associated with better functional neuroimaging outcomes, such as enhanced frontal, parietal, and occipital activation and resting-state functional connectivity [120]. Environmental enrichment has been linked to enhanced plasticity and cortical remodelling through microglial engulfment of PNNs; socially-driven oxytocin boosts; and secretion of dopamine, serotonin, and noradrenaline [141]. Thus, these interventions may reverse excessive synaptic loss and/or prevent further deterioration. Interventions targeting stress and well-being have also been investigated, with integrated coping awareness therapy, mindfulness-based stress reduction, and cognitive-behavioural therapy showing promising results [303–305]. However, investigations on the neural and cognitive changes resulting from these therapies are rare. One such trial reported that differences in brain activations between patients and controls were ameliorated by CBT targeting 'jumping to conclusions' and attributional biases, a prominent cognitive style in patients that can produce delusions [306]. Additionally, whether the aforementioned interventions mitigate excessive peri-adolescent SP is unknown. Longitudinal studies should follow a sample of healthy children, children showing early signs of psychosis without treatment, and children with early signs of psychosis receiving the aforementioned therapies into late adolescence and compare the structural, functional, and cognitive outcomes of these three groups.

IV. CONCLUSION

This paper narrates the literature on a variety of factors and mechanisms related to SP to various degrees of closeness. We aspired to delineate the broad span of this hypothesis and thereby the ideas it offers to future investigations. SCZ is a complex disorder with several evidence-based theories explaining it, but no single unified framework has been developed to date. Thanks to its diffuse nature, SP seems to be a

point of convergence for some of the aetiologic mechanisms proposed so far, namely genes, immune dysfunction, neuroinflammation, neural dysconnectivity, compromised plasticity, hyper- and hypo-glutamatergic states, and environmental influences. However, the same diffuse nature also allows it to overlap with competing mechanisms that may manifest similarly, such as deficient synaptogenesis, high synaptic apoptosis, excitotoxicity, and enfeebled cytoarchitecture. It is important to note that SP cannot yet be directly observed *in vivo*; our current understanding relies largely on indirect measures from neuroimaging and postmortem studies. Nonetheless, the developmental basis of SP gives us hope that attempts to correct it in children at risk or showing early signs of psychosis may lead to a less severe and debilitating form of SCZ. For SP to become a reliable target of interventions, future research would need to disentangle it from competing mechanisms and fortify its importance in SCZ by investigating it through a variety of tools and designs, especially longitudinal studies.

V. REFERENCES

1. Boland R, Verduin ML, editors. Kaplan and Sadock's comprehensive textbook of psychiatry. 11th ed. Philadelphia: Lippincott Williams & Wilkins; 2024.
2. Feinberg I. Schizophrenia: caused by a fault in programmed synaptic elimination during adolescence? *J Psychiatr Res.* 1982;17(4):319–34.
3. Faust TE, Gunner G, Schafer DP. Mechanisms governing activity-dependent synaptic pruning in the developing mammalian CNS. *Nat Rev Neurosci.* 2021;22(11):657–73.
4. Mallya AP, Wang HD, Lee HNR, Deutch AY. Microglial pruning of synapses in the prefrontal cortex during adolescence. *Cereb Cortex.* 2019;29(4):1634–1643.
5. Riccomagno MM, Kolodkin AL. Sculpting neural circuits by axon and dendrite pruning. *Annu Rev Cell Dev Biol.* 2015;31:779–805.
6. Koyama R, Ikegaya Y. Microglia in the pathogenesis of autism spectrum disorders. *Neurosci Res.* 2015;100:1–5.
7. Pardo CA, Vargas DL, Zimmerman AW. Immunity, neuroglia and neuroinflammation in autism. *Int Rev Psychiatry.* 2005;17:485–495.
8. Marco C, Antonio D, Antonina S, Alessandro S, Concetta C. Genes involved in pruning and inflammation are enriched in a large mega-sample of patients affected by schizophrenia and bipolar disorder and controls. *Psychiatry Res.* 2015;228(3):945–949.
9. Peca J, Feng G. Cellular and synaptic network defects in autism. *Curr Opin Neurobiol.* 2012;22:866–872.
10. Thomas MS, Davis R, Karmiloff-Smith A, Knowland VC, Charman T. The over-pruning hypothesis of autism. *Dev Sci.* 2016;19:284–305.
11. Waites CL, Garner CC. Presynaptic function in health and disease. *Trends Neurosci.* 2011;34:326–337.
12. Melom JE, Littleton JT. Synapse development in health and disease. *Curr Opin Genet Dev.* 2011;21:256–261.
13. Penzes P, Cahill ME, Jones KA, VanLeeuwen JE, Woolfrey KM. Dendritic spine pathology in neuropsychiatric disorders. *Nat Neurosci.* 2011;14:285–293.
14. Zhang MM, Guo MX, Zhang QP, Chen XQ, Li NZ, Liu Q, et al. IL-1R/C3aR signaling regulates synaptic pruning in the prefrontal cortex of depression. *Cell Biosci.* 2022;12(1):90.

15. American Psychiatric Association. Diagnostic and statistical manual of mental disorders. 5th ed., text rev. Washington, DC: APA; 2022. Available from: <https://doi.org/10.1176/appi.books.9780890425787>
16. Lawrie SM, Abukmeil SS. Brain abnormality in schizophrenia: A systematic and quantitative review of volumetric magnetic resonance imaging studies. *Br J Psychiatry*. 1998;172:110–20.
17. Steen RG, Mull C, McClure R, Hamer RM, Lieberman JA. Brain volume in first-episode schizophrenia: systematic review and meta-analysis of magnetic resonance imaging studies. *Br J Psychiatry*. 2006;188:510–518.
18. Levitt JJ, Bobrow L, Lucia D, Srinivasan P. A selective review of volumetric and morphometric imaging in schizophrenia. *Behavioral neurobiology of schizophrenia and its treatment*. 2010:243–281.
19. Van Haren NE, Cahn W, Pol HH, Kahn RS. Confounders of excessive brain volume loss in schizophrenia. *Neurosci Biobehav Rev*. 2013;37(10):2418–2423.
20. Kuo SS, Pogue-Geile MF. Variation in fourteen brain structure volumes in schizophrenia: A comprehensive meta-analysis of 246 studies. *Neurosci Biobehav Rev*. 2019;98:85–94.
21. Haijma SV, Van Haren N, Cahn W, Koolschijn PC, Hulshoff Pol HE, Kahn RS. Brain volumes in schizophrenia: a meta-analysis in over 18,000 subjects. *Schizophr Bull*. 2013;39(5):1129–38.
22. McIntosh AM, Owens DC, Moorhead WJ, Whalley HC, Stanfield AC, Hall J, et al. Longitudinal volume reductions in people at high genetic risk of schizophrenia as they develop psychosis. *Biol Psychiatry*. 2011;69:953–958.
23. Harrison PJ. The hippocampus in schizophrenia: a review of the neuropathological evidence and its pathophysiological implications. *Psychopharmacology (Berl)*. 2004;174:151–62.
24. Tamminga CA, Stan AD, Wagner AD. The hippocampal formation in schizophrenia. *Am J Psychiatry*. 2010;167:1178–1193.
25. Bogerts B, Meertz E, Schönfeldt-Bausch R. Basal ganglia and limbic system pathology in schizophrenia. A morphometric study of brain volume and shrinkage. *Arch Gen Psychiatry*. 1985;42:784–91.
26. Brandt GN, Bonelli RM. Structural neuroimaging of the basal ganglia in schizophrenic patients: a review. *Wien Med Wochenschr*. 2008;158:84–90.
27. Horga G, Bernacer J, Dusi N, Entis J, Chu K, Hazlett EA, et al. Correlations between ventricular enlargement and gray and white matter volumes of cortex, thalamus, striatum, and internal capsule in schizophrenia. *Eur Arch Psychiatry Clin Neurosci*. 2011;261:467–76.
28. Supprian T, Ulmar G, Bauer M, Schuler M, Puschel K, Retz-Junginger P, Schmitt HP, Heinsen H. Cerebellar vermis area in schizophrenic patients—a post-mortem study. *Schizophr Res*. 2000;42:19–28.
29. Andersen BB, Pakkenberg B. Stereological quantitation in cerebella from people with schizophrenia. *Br J Psychiatry*. 2003;182:354–61.
30. Kakeda S, Korogi Y. The efficacy of a voxel-based morphometry on the analysis of imaging in schizophrenia, temporal lobe epilepsy, and Alzheimer's disease/mild cognitive impairment: a review. *Neuroradiology*. 2010;52:711–21.
31. Glahn DC, Laird AR, Ellison-Wright I, Thelen SM, Robinson JL, Lancaster JL, et al. Meta-analysis of gray matter anomalies in schizophrenia: application of anatomic likelihood estimation and network analysis. *Biol Psychiatry*. 2008;64(9):774–81.

32. Rimol LM, Nesvåg R, Hagler DJ Jr, Bergmann Ø, Fennema-Notestine C, Hartberg CB, et al. Cortical volume, surface area, and thickness in schizophrenia and bipolar disorder. *Biol Psychiatry*. 2012;71(6):552–60.
33. Marsh R, Gerber AJ, Peterson BS. Neuroimaging studies of normal brain development and their relevance for understanding childhood neuropsychiatric disorders. *Journal of the American Academy of Child & Adolescent Psychiatry*. 2008 Nov 1;47(11):1233-51.
34. Sowell ER, Thompson PM, Tessner KD, Toga AW. Mapping continued brain growth and gray matter density reduction in dorsal frontal cortex: Inverse relationships during postadolescent brain maturation. *Journal of Neuroscience*. 2001 Nov 15;21(22):8819-29.
35. Fusar-Poli P, Broome MR, Woolley JB, Johns LC, Tabraham P, Bramon E, Valmaggia L, Williams SC, McGuire P. Altered brain function directly related to structural abnormalities in people at ultra high risk of psychosis: longitudinal VBM-fMRI study. *Journal of psychiatric research*. 2011 Feb 1;45(2):190-8.
36. Cannon TD, Thompson PM, van Erp TGM, Toga AW, Poutanen V-P, Huttunen M, et al. Cortex mapping reveals regionally specific patterns of genetic and disease-specific gray-matter deficits in twins discordant for schizophrenia. *Proc Natl Acad Sci U S A*. 2002;99:3228–33.
37. Hulshoff Pol HE, Brans RG, van Haren NE, Schnack HG, Langen M, Baare WF, van Oel CJ, Kahn RS. Gray and white matter volume abnormalities in monozygotic and same-gender dizygotic twins discordant for schizophrenia. *Biol Psychiatry*. 2004;55:126–130.
38. Borgwardt SJ, Picchioni MM, Ettinger U, Touloupoulou T, Murray R, McGuire PK. Regional gray matter volume in monozygotic twins concordant and discordant for schizophrenia. *Biol Psychiatry*. 2010;67:956–64.
39. Mechelli A, Riecher-Rossler A, Meisenzahl EM, Tognin S, Wood SJ, Borgwardt SJ, et al. Neuroanatomical abnormalities that predate the onset of psychosis: a multicenter study. *Arch Gen Psychiatry*. 2011;68:489–495.
40. Andreasen NC, Nopoulos P, Magnotta V, Pierson R, Ziebell S, Ho BC. Progressive brain change in schizophrenia: a prospective longitudinal study of first-episode schizophrenia. *Biological psychiatry*. 2011 Oct 1;70(7):672-9.
41. Chwa WJ, Tishler TA, Raymond C, Tran C, Anwar F, Villablanca JP, et al. Association between cortical volume and gray-white matter contrast with second generation antipsychotic medication exposure in first episode male schizophrenia patients. *Schizophr Res*. 2020;222:397–410.
42. Sun X, Yue S, Duan M, Yao D, Luo C. Psychosocial intervention for schizophrenia. *Brain-Apparatus Commun*. 2023;2(1):2178266.
43. Takahashi T, Wood SJ, Yung AR, Soulsby B, McGorry PD, Suzuki M, Kawasaki Y, Phillips LJ, Velakoulis D, Pantelis C. Progressive gray matter reduction of the superior temporal gyrus during transition to psychosis. *Arch Gen Psychiatry*. 2009;66:366–376.
44. Hazlett EA, Buchsbaum MS, Haznedar MM, Newmark R, Goldstein KE, Zelmanova Y, Glanton CF, Torosjan Y, New AS, Lo JN, Mitropoulou V. Cortical gray and white matter volume in unmedicated schizotypal and schizophrenia patients. *Schizophrenia research*. 2008 Apr 1;101(1-3):111-23.
45. Honea R, Crow TJ, Passingham D, Mackay CE. Regional deficits in brain volume in schizophrenia: a meta-analysis of voxel-based morphometry studies. *Am J Psychiatry*. 2005;162:2233–45.

46. Okugawa G, Sedvall GC, Agartz I. Reduced grey and white matter volumes in the temporal lobe of male patients with chronic schizophrenia. *European archives of psychiatry and clinical neuroscience*. 2002 Jun;252:120-3.
47. Okugawa G, Tamagaki C, Agartz I. Frontal and temporal volume size of grey and white matter in patients with schizophrenia: an MRI parcellation study. *European archives of psychiatry and clinical neuroscience*. 2007 Aug;257:304-7.
48. Nygård M, Løberg EM, Craven AR, Ersland L, Berle JØ, Kroken RA, et al. Dichotic listening, executive functions and grey matter cortical volume in patients with schizophrenia and healthy controls. *Scand J Psychol*. 2013;54(6):443-450.
49. Zhang Y, Catts VS, Sheedy D, McCrossin T, Kril JJ, Shannon Weickert C. Cortical grey matter volume reduction in people with schizophrenia is associated with neuro-inflammation. *Transl Psychiatry*. 2016;6(12):e982-e982.
50. Thompson PM, Vidal C, Giedd JN, Gochman P, Blumenthal J, Nicolson R, et al. Mapping adolescent brain change reveals dynamic wave of accelerated gray matter loss in very early-onset schizophrenia. *Proc Natl Acad Sci*. 2001;98(20):11650-11655.
51. Premkumar P, Fannon D, Kuipers E, Cooke MA, Simmons A, Kumari V. Association between a longer duration of illness, age and lower frontal lobe grey matter volume in schizophrenia. *Behavioural Brain Research*. 2008 Nov 3;193(1):132-9.
52. Kim GW, Kim YH, Jeong GW. Whole brain volume changes and its correlation with clinical symptom severity in patients with schizophrenia: A DARTEL-based VBM study. *PLoS One*. 2017;12(5):e0177251.
53. Wannan CM, Croypley VL, Chakravarty MM, Bousman C, Ganella EP, Bruggemann JM, Weickert TW, Weickert CS, Everall I, McGorry P, Velakoulis D. Evidence for network-based cortical thickness reductions in schizophrenia. *American Journal of Psychiatry*. 2019 Jul 1;176(7):552-63.
54. Malwade S, Lekander JG, Samudiyata S, Koskivi M, Storvik M, Sellgren CM. Elevated synaptic pruning by microglia in schizophrenia across patient-derived brain organoids. *Neuroscience Applied*. 2023 Jan 1;2:102643.
55. Shenton ME, Whitford TJ, Kubicki M. Structural neuroimaging in schizophrenia from methods to insights to treatments. *Dialogues Clin Neurosci*. 2010;12(3):317-32.
56. Wheeler AL, Voineskos AN. A review of structural neuroimaging in schizophrenia: from connectivity to connectomics. *Front Hum Neurosci*. 2014;8:653.
57. Honea R, Crow TJ, Passingham D, Mackay CE. Regional deficits in brain volume in schizophrenia: a meta-analysis of voxel-based morphometry studies. *Am J Psychiatry*. 2005;162:2233-45.
58. Jorgensen KN, Nerland S, Norbom LB, Doan NT, Nesvag R, Mørch-Johnsen L, et al. Increased MRI-based cortical grey/white-matter contrast in sensory and motor regions in schizophrenia and bipolar disorder. *Psychol Med*. 2016;46(9):1971-85.
59. Buyanova IS, Arsalidou M. Cerebral white matter myelination and relations to age, gender, and cognition: a selective review. *Front Hum Neurosci*. 2021;15:662031.
60. Huttenlocher PR. *Neural Plasticity: The Effects of the Environment on the Development of the Cerebral Cortex*. Cambridge: Harvard University Press; 2002.
61. Najjar S, Pearlman DM. Neuroinflammation and white matter pathology in schizophrenia: systematic review. *Schizophr Res*. 2015;161(1):102-112.

62. Glausier JR, Lewis DA. Dendritic spine pathology in schizophrenia. *Neuroscience*. 2013;251:90–107.
63. Thune JJ, Uylings HB, Pakkenberg B. No deficit in total number of neurons in the prefrontal cortex in schizophrenics. *J Psychiatr Res*. 2001;35(1):15-21. [http://dx.doi.org/10.1016/S0022-3956\(00\)00043-1](http://dx.doi.org/10.1016/S0022-3956(00)00043-1)
64. Selemon LD, Rajkowska G, Goldman-Rakic PS. Abnormally high neuronal density in the schizophrenic cortex. A morphometric analysis of prefrontal area 9 and occipital area 17. *Arch Gen Psychiatry*. 1995;52(10):805–18.
65. Selemon LD, Rajkowska G, Goldman-Rakic PS. Elevated neuronal density in prefrontal area 46 in brains from schizophrenic patients: application of a three-dimensional, stereologic counting method. *J Comp Neurol*. 1998;392(3):402–12.
66. DeFelipe J, Fariñas I. The pyramidal neuron of the cerebral cortex: morphological and chemical characteristics of the synaptic inputs. *Progress in neurobiology*. 1992 Dec 1;39(6):563-607.
67. Rochefort NL, Konnerth A. Dendritic spines: from structure to in vivo function. *EMBO Rep*. 2012;13(8):699–708.
68. Wilson CJ. GABAergic inhibition in the neostriatum. *Progress in brain research*. 2007 Jan 1;160:91-110.
69. Nimchinsky EA, Sabatini BL, Svoboda K. Structure and function of dendritic spines. *Annu Rev Physiol*. 2002;64(1):313-353.
70. Yuste R. Dendritic spines and distributed circuits. *Neuron*. 2011 Sep 8;71(5):772-81.
71. Berdenis van Berlekom A, Mufliah CH, Snijders GJ, MacGillavry HD, Middeldorp J, Hol EM, et al. Synapse pathology in schizophrenia: a meta-analysis of postsynaptic elements in postmortem brain studies. *Schizophr Bull*. 2020;46(2):374–86.
72. Moyer CE, Shelton MA, Sweet RA. Dendritic spine alterations in schizophrenia. *Neuroscience letters*. 2015 Aug 5;601:46-53.
73. Glantz LA, Lewis DA. Decreased dendritic spine density on prefrontal cortical pyramidal neurons in schizophrenia. *Arch Gen Psychiatry*. 2000;57(1):65–73.
74. Roberts RC, Conley R, Kung L, Peretti FJ, Chute DJ. Reduced striatal spine size in schizophrenia: a postmortem ultrastructural study. *Neuroreport*. 1996;7(6):1214–8.
75. Rosoklija G, Toomayan G, Ellis SP, Keilp J, Mann JJ, Latov N, et al. Structural abnormalities of subicular dendrites in subjects with schizophrenia and mood disorders: Preliminary findings. *Arch Gen Psychiatry*. 2000;57(4):349–56.
76. Garey LJ, Ong WY, Patel TS, Kanani M, Davis AM, Mortimer AM, et al. Reduced dendritic spine density on cerebral cortical pyramidal neurons in schizophrenia. *J Neurol Neurosurg Psychiatry*. 1998;65(4):446–53. <https://doi.org/10.1136/jnnp.65.4.446>
77. Sweet RA, Henteleff RA, Zhang W, Sampson AR, Lewis DA. Reduced dendritic spine density in auditory cortex of subjects with schizophrenia. *Neuropsychopharmacology*. 2009;34(4):374-389.
78. Konopaske GT, Lange N, Coyle JT, Benes FM. Prefrontal cortical dendritic spine pathology in schizophrenia and bipolar disorder. *JAMA Psychiatry*. 2014;71(12):1323–31.
79. Dienel, S. J., Schoonover, K. E., & Lewis, D. A. (2022). Cognitive dysfunction and prefrontal cortical circuit alterations in schizophrenia: developmental trajectories. *Biological psychiatry*, 92(6), 450-459.

80. Liston C, Miller MM, Goldwater DS, Radley JJ, Rocher AB, Hof PR, et al. Stress-induced alterations in prefrontal cortical dendritic morphology predict selective impairments in perceptual attentional set-shifting. *J Neurosci*. 2006;26(30):7870-7874.
81. Hains AB, Vu MAT, Maciejewski PK, van Dyck CH, Gottron M, Arnsten AFT. Inhibition of protein kinase C signaling protects prefrontal cortex dendritic spines and cognition from the effects of chronic stress. *Proc Natl Acad Sci USA*. 2009;106(42):17957–62.
82. Parellada E, Gassó P. Glutamate and microglia activation as a driver of dendritic apoptosis: a core pathophysiological mechanism to understand schizophrenia. *Translational psychiatry*. 2021 May 6;11(1):271.
83. Chen CC, Lu J, Zuo Y. Spatiotemporal dynamics of dendritic spines in the living brain. *Front Neuroanat*. 2014;8:28.
84. Chung WS, Welsh CA, Barres BA, Stevens B. Do glia drive synaptic and cognitive impairment in disease? *Nat Neurosci*. 2015;18(11):1539–45.
85. Chen X, Sun C, Chen Q, et al. Apoptotic engulfment pathway and schizophrenia. *PLoS One*. 2009;4(9):e6875.
86. Sellgren CM, Gracias J, Watmuff B, Biag JD, Thanos JM, Whittredge PB, et al. Increased synapse elimination by microglia in schizophrenia patient-derived models of synaptic pruning. *Nat Neurosci*. 2019;22(3):374–85.
87. Forrest MP, Parnell E, Penzes P. Dendritic structural plasticity and neuropsychiatric disease. *Nat Rev Neurosci*. 2018;19(4):215–34.
88. Holtmaat AJ, Trachtenberg JT, Wilbrecht L, Shepherd GM, Zhang X, Knott GW, et al. Transient and persistent dendritic spines in the neocortex in vivo. *Neuron*. 2005;45(2):279–91.
89. Ofer N, Berger DR, Kasthuri N, Lichtman JW, Yuste R. Ultrastructural analysis of dendritic spine necks reveals a continuum of spine morphologies. *Dev Neurobiol*. 2021;81(5):746–757.
90. Arnsten AF, Paspalas CD, Gamo NJ, Yang Y, Wang M. Dynamic Network Connectivity: A new form of neuroplasticity. *Trends Cogn Sci*. 2010;14:365–75.
91. Paulin JJW, Haslehurst P, Fellows AD, Liu W, Jackson JD, Joel Z, et al. Large and small dendritic spines serve different interacting functions in hippocampal synaptic plasticity and homeostasis. *Neural Plast*. <https://doi.org/10.1155/2016/6170509>
92. Bourne J, Harris KM. Do thin spines learn to be mushroom spines that remember? *Curr Opin Neurobiol*. 2007;17:381–6.
93. Kasai H, Matsuzaki M, Noguchi J, Yasumatsu N, Nakahara H. Structure-stability-function relationships of dendritic spines. *Trends Neurosci*. 2003;26:360–8.
94. Zuo Y, Lin A, Chang P, Gan WB. Development of long-term dendritic spine stability in diverse regions of cerebral cortex. *Neuron*. 2005 Apr 21;46(2):181-9.
95. Xu T, Yu X, Perlik AJ, Tobin WF, Zweig JA, Tennant K, et al. Rapid formation and selective stabilization of synapses for enduring motor memories. *Nature*. 2009;462:915–919.
96. Holtmaat A, Svoboda K. Experience-dependent structural synaptic plasticity in the mammalian brain. *Nat Rev Neurosci*. 2009;10:647–58.
97. MacDonald ML, Alhassan J, Newman JT, Richard M, Gu H, Kelly RM, et al. Selective loss of smaller spines in schizophrenia. *Am J Psychiatry*. 2017;174(6):586-594.

98. McKinney BC, MacDonald ML, Newman JT, Shelton MA, DeGiosio RA, Kelly RM, et al. Density of small dendritic spines and microtubule-associated-protein-2 immunoreactivity in the primary auditory cortex of subjects with schizophrenia. *Neuropsychopharmacology*. 2019;44(6):1055-1061.
99. Cai X, Yang ZH, Li HJ, Xiao X, Li M, Chang H. A human-specific schizophrenia risk tandem repeat affects alternative splicing of a human-unique isoform AS3MT d2d3 and mushroom dendritic spine density. *Schizophr Bull*. 2021;47(1):219–27.
100. Sánchez-González A, Thougard E, Tapias-Espinosa C, Cañete T, Sampedro-Viana D, Saunders JM, et al. Increased thin-spine density in frontal cortex pyramidal neurons in a genetic rat model of schizophrenia-relevant features. *Eur Neuropsychopharmacol*. 2021;44:79–91.
101. Gonzalez-Burgos G, Miyamae T, Nishihata Y, Krimer OL, Lewis DA. Strength of excitatory inputs to layer 3 pyramidal neurons during synaptic pruning in the monkey prefrontal cortex: Relevance for the pathogenesis of schizophrenia. *Biological psychiatry*. 2023 Aug 15;94(4):288-96.
102. Heinrichs RW, Zakzanis KK. Neurocognitive deficit in schizophrenia: a quantitative review of the evidence. *Neuropsychology*. 1998;12:426–45.
103. Häfner H, Riecher-Rössler A, Hambrecht M, Maurer K, Meissner S, Schmidtke A, et al. IRAOS: an instrument for the assessment of onset and early course of schizophrenia. *Schizophr Res*. 1992;6:209–23.
104. Rund BR. A review of longitudinal studies of cognitive functions in schizophrenia patients. *Schizophr Bull*. 1998;24:425–35.
105. Bhandari A, Voineskos D, Daskalakis ZJ, Rajji TK, Blumberger DM. A review of impaired neuroplasticity in schizophrenia investigated with non-invasive brain stimulation. *Front Psychiatry*. 2016;7:45.
106. Tripathi A, Kar SK, Shukla R. Cognitive Deficits in Schizophrenia: Understanding the Biological Correlates and Remediation Strategies. *Clin Psychopharmacol Neurosci*. 2018;16(1):7-17. doi: 10.9758/cpn.2018.16.1.7.
107. Voineskos D, Rogasch NC, Rajji TK, Fitzgerald PB, Daskalakis ZJ. A review of evidence linking disrupted neural plasticity to schizophrenia. *Can J Psychiatry*. 2013;58:86–92. doi: 10.1177/070674371305800205.
108. Van Snellenberg JX. Working memory and long-term memory deficits in schizophrenia: is there a common substrate? *Psychiatry Res*. 2009;174:89-96.
109. Daskalakis ZJ, Christensen BK, Fitzgerald PB, Chen R. Dysfunctional neural plasticity in patients with schizophrenia. *Arch Gen Psychiatry*. 2008;65:378–85.
110. Mahdavi A, Bahrami F, Janahmadi M. Analysis of impaired LTP in schizophrenia using an extended mathematical model of a tripartite synapse. In: 2015 22nd Iranian Conference on Biomedical Engineering (ICBME). IEEE; 2015. p. 76-80.
111. Forsyth JK, Lewis DA. Mapping the consequences of impaired synaptic plasticity in schizophrenia through development: an integrative model for diverse clinical features. *Trends Cogn Sci*. 2017;21(10):760–78.
112. XL, Yan QJ, Zhu F. Abnormal synaptic plasticity and impaired cognition in schizophrenia. *World J Psychiatry*. 2022;12(4):541.
113. Ripke S, Neale BM, Corvin A, Walters JT, Farh KH, Holmans PA, Lee P, Bulik-Sullivan B, Collier DA, Huang HL, Pers TH. Biological insights from 108 schizophrenia-associated genetic loci. *Nature*. 2014;511(7510):421-7. <https://doi.org/10.1038/nature13595>

114. Li S, Hu N, Zhang W, Tao B, Dai J, Gong Y, et al. Dysconnectivity of multiple brain networks in schizophrenia: a meta-analysis of resting-state functional connectivity. *Front Psychiatry*. 2019;10:482.
115. Van Den Heuvel MP, Fornito A. Brain networks in schizophrenia. *Neuropsychol Rev*. 2014;24:32-48.
116. Barlati S, Nibbio G, Vita A. Evidence-based psychosocial interventions in schizophrenia: a critical review. *Curr Opin Psychiatry*. 2024;37(3):131–9.
117. Kar SK, Singh A. Neuroplasticity and cognitive training in schizophrenia. *Curr Behav Neurosci Rep*. 2019;6:113–18.
118. Thorsen AL, Johansson K, Løberg EM. Neurobiology of cognitive remediation therapy for schizophrenia: a systematic review. *Front Psychiatry*. 2014;5:103.
119. Solmi M, Croatto G, Piva G, Rosson S, Fusar-Poli P, Rubio JM, et al. Efficacy and acceptability of psychosocial interventions in schizophrenia: systematic overview and quality appraisal of the meta-analytic evidence. *Mol Psychiatry*. 2023;28(1):354–68.
120. Matsuda Y, Makinodan M, Morimoto T, Kishimoto T. Neural changes following cognitive remediation therapy for schizophrenia. *Psychiatry Clin Neurosci*. 2019;73(11):676-684.
121. Nägerl UV, Eberhorn N, Cambridge SB, Bonhoeffer T. Bidirectional activity-dependent morphological plasticity in hippocampal neurons. *Neuron*. 2004;44(5):759-767. <https://doi.org/10.1016/j.neuron.2004.11.016>
122. De Roo M, Klauser P, Muller D. LTP promotes a selective long-term stabilization and clustering of dendritic spines. *PLoS Biol*. 2008;6(9):e219. <https://doi.org/10.1371/journal.pbio.0060219>
123. Zhou Q, Homma KJ, Poo MM. Shrinkage of dendritic spines associated with long-term depression of hippocampal synapses. *Neuron*. 2004;44(5):749-757. doi:10.1016/j.neuron.2004.11.011.
124. Bastrikova N, Gardner GA, Reece JM, Jeromin A, Dudek SM. Synapse elimination accompanies functional plasticity in hippocampal neurons. *Proc Natl Acad Sci U S A*. 2008;105(8):3123–7.
125. Becker N, Wierenga CJ, Fonseca R, Bonhoeffer T, Nägerl UV. LTD induction causes morphological changes of presynaptic boutons and reduces their contacts with spines. *Neuron*. 2008;60(4):590–7.
126. Malenka RC, Bear MF. LTP and LTD: An embarrassment of riches. *Neuron*. 2004;44(1):5–21. <https://doi.org/10.1016/j.neuron.2004.09.012>
127. Bliss TV, Collingridge GL. A synaptic model of memory: Long-term potentiation in the hippocampus. *Nature*. 1993;361(6407):31–9.
128. Winterer G, et al. Prefrontal broadband noise, working memory and genetic risk for schizophrenia. *Am J Psychiatry*. 2004;161:490–500.
129. Hasan A, Nitsche MA, Rein B, Schneider-Axmann T, Guse B, Gruber O, et al. Dysfunctional long-term potentiation-like plasticity in schizophrenia revealed by transcranial direct current stimulation. *Behav Brain Res*. 2011;224(1):15–22.
130. Strube W, Bunse T, Nitsche MA, Wobrock T, Aborowa R, Misewitsch K, et al. Smoking restores impaired LTD-like plasticity in schizophrenia: a transcranial direct current stimulation study. *Neuropsychopharmacology*. 2015;40(4):822-830.
131. Frantseva MV, Fitzgerald PB, Chen R, Möller B, Daigle M, Daskalakis ZJ. Evidence for impaired long-term potentiation in schizophrenia and its relationship to motor skill learning. *Cereb Cortex*. 2008;18(5):990–96.

132. Valstad M, Roelfs D, Slapø NB, Timpe CM, Rai A, Matziorinis AM, et al. Evidence for reduced long-term potentiation-like visual cortical plasticity in schizophrenia and bipolar disorder. *Schizophr Bull.* 2021;47(6):1751-1760.
133. Palm U, Lustig D, Frick B, Nitsche MA, Kuo MF, Jobst A, et al. Transcranial direct currents stimulation reveals overshooting LTP-like motor cortex plasticity in schizophrenia: A study in antipsychotic-free patients.
134. Labrou D. Dopamine D5 receptor involvement in LTP and LTD: adjustment to the dysconnectivity theory of schizophrenia. *Qeios.* 2022.
135. Lewis DA, Glantz LA, Pierri JN, Sweet RA. Altered cortical glutamate neurotransmission in schizophrenia: evidence from morphological studies of pyramidal neurons. *Ann N Y Acad Sci.* 2003;1003:102-112.
136. Paz RD, Tardito S, Atzori M, Tseng KY. Glutamatergic dysfunction in schizophrenia: from basic neuroscience to clinical psychopharmacology. *Eur Neuropsychopharmacol.* 2008;18:773-786.
137. Woo TU, Shrestha K, Lamb D, Minns MM, Benes FM. N-methyl-D-aspartate receptor and calbindin-containing neurons in the anterior cingulate cortex in schizophrenia and bipolar disorder. *Biol Psychiatry.* 2008;64:803-809.
138. Woo TU, Walsh JP, Benes FM. Density of glutamic acid decarboxylase 67 messenger RNA containing neurons that express the N-methyl-D-aspartate receptor subunit NR2A in the anterior cingulate cortex in schizophrenia and bipolar disorder. *Arch Gen Psychiatry.* 2004;61:649-657.
139. Yonezawa K, Tani H, Nakajima S, Nagai N, Koizumi T, Miyazaki T, et al. AMPA receptors in schizophrenia: A systematic review of postmortem studies on receptor subunit expression and binding. *Schizophr Res.* 2022;243:98-109.
140. Hasan A, Nitsche MA, Herrmann M, Schneider-Axmann T, Marshall L, Gruber O, et al. Impaired long-term depression in schizophrenia: a cathodal tDCS pilot study. *Brain Stimul.* 2012;5(4):475–83.
141. Tooley UA, Bassett DS, Mackey AP. Environmental influences on the pace of brain development. *Nat Rev Neurosci.* 2021;22(6):372-384.
142. Kuznetsov VV, Kruchinin EV, Aliev FS, Autlev KM, Yanin EL. Molecular mechanisms of synaptic pruning regulation. *J Crit Rev.* 2020;7(12):515–8.
143. Scholl C, Rule ME, Hennig MH. The information theory of developmental pruning: Optimizing global network architectures using local synaptic rules. *PLoS Comput Biol.* 2021;17(10):e1009458.
144. Millána AP, Torresa JJ, Johnsonb S, Marroa J. The concurrence of form and function in developing networks: An explanation for synaptic pruning. *arXiv preprint.* 2017;arXiv:1705.02773.
145. Zhao F, Zeng Y. Dynamically optimizing network structure based on synaptic pruning in the brain. *Front Syst Neurosci.* 2021;15:620558.
146. Stephan KE, Friston KJ, Frith CD. Dysconnection in schizophrenia: from abnormal synaptic plasticity to failures of self-monitoring. *Schizophr Bull.* 2009;35(3):509–27. doi:10.1093/schbul/sbn176.
147. Winterer G, Weinberger DR. Genes, dopamine and cortical signal-to-noise ratio in schizophrenia. *Trends Neurosci.* 2004;27(11):683-690.
148. Arkhipov AY, Nurbekov MK, Strelets VB. Incongruence of neurophysiological manifestations as reflection of disturbances in synaptic pruning of the cortex in paranoid schizophrenia.

149. Khandaker GM, Cousins L, Deakin J, Lennox BR, Yolken R, Jones PB. Inflammation and immunity in schizophrenia: implications for pathophysiology and treatment. *The Lancet Psychiatry*. 2015 Mar 1;2(3):258-70.
150. Miller BJ, et al. Inflammation and schizophrenia: a review. *Schizophr Bull*. 2011;37(5):1020-1030.
151. Benros ME, Nielsen PR, Nordentoft M, Eaton WW, Dalton SO, Mortensen PB. Autoimmune diseases and severe infections as risk factors for schizophrenia: a 30-year population-based register study. *American Journal of Psychiatry*. 2011 Dec;168(12):1303-10.
152. Stephan AH, Barres BA, Stevens B. The complement system: an unexpected role in synaptic pruning during development and disease. *Annual review of neuroscience*. 2012 Jul 21;35(1):369-89.
153. Muller N, J. Schwarz M. The role of immune system in schizophrenia. *Current immunology reviews*. 2010 Aug 1;6(3):213-20.
154. Meyer U, Weiner I, McAlonan GM, Feldon J. The neuropathological contribution of prenatal inflammation to schizophrenia. *Expert review of neurotherapeutics*. 2011 Jan 1;11(1):29-32.
155. Saetre P, Emilsson L, Axelsson E, Kreuger J, Lindholm E, Jazin E. Inflammation-related genes up-regulated in schizophrenia brains. *Bmc Psychiatry*. 2007 Dec;7:1-0.
156. Zhang Y, Catts VS, Sheedy D, McCrossin T, Kril JJ, Shannon Weickert C. Cortical grey matter volume reduction in people with schizophrenia is associated with neuro-inflammation. *Transl Psychiatry*. 2016;6(12):e982-e982.
157. Woodburn SC, Bollinger JL, Wohleb ES. Synaptic and behavioral effects of chronic stress are linked to dynamic and sex-specific changes in microglia function and astrocyte dystrophy. *Neurobiology of stress*. 2021 May 1;14:100312.
158. Bayer TA, Buslei R, Havas L, Falkai P. Evidence for activation of microglia in patients with psychiatric illnesses. *Neuroscience letters*. 1999 Aug 20;271(2):126-8.
159. Radewicz K, Garey LJ, Gentleman SM, Reynolds R. Increase in HLA-DR immunoreactive microglia in frontal and temporal cortex of chronic schizophrenics. *Journal of Neuropathology & Experimental Neurology*. 2000 Feb 1;59(2):137-50.
160. van Kesteren CFMG, Gremmels H, de Witte LD, et al. Immune involvement in the pathogenesis of schizophrenia: a meta-analysis on postmortem brain studies. *Transl Psychiatry*. 2017;7:e1075.
161. Trubetskoy V, Pardiñas AF, Qi T, Panagiotaropoulou G, Awasthi S, Bigdeli TB, Bryois J, Chen CY, Dennison CA, Hall LS, Lam M. Mapping genomic loci implicates genes and synaptic biology in schizophrenia. *Nature*. 2022 Apr 21;604(7906):502-8.
162. Sekar A, Bialas AR, De Rivera H, Davis A, Hammond TR, Kamitaki N, Tooley K, Presumey J, Baum M, Van Doren V, Genovese G. Schizophrenia risk from complex variation of complement component 4. *Nature*. 2016 Feb 11;530(7589):177-83.
163. Yilmaz M, Yalcin E, Presumey J, Aw E, Ma M, Whelan CW, Stevens B, McCarroll SA, Carroll MC. Overexpression of schizophrenia susceptibility factor human complement C4A promotes excessive synaptic loss and behavioral changes in mice. *Nature neuroscience*. 2021 Feb;24(2):214-24.
164. Stevens B, Allen NJ, Vazquez LE, Howell GR, Christopherson KS, Nouri N, et al. The classical complement cascade mediates CNS synapse elimination. *Cell*. 2007;131:1164–1178.
165. Sheridan SD, Horng JE, Yeh H, McCrea L, Fu T, Perlis RH. Loss of function in the autism and schizophrenia-associated gene CYFIP1 in human microglia supports a role in synaptic pruning. *bioRxiv*. 2022 Oct 24:2022-10.

166. Yilmaz M. Role of Schizophrenia Risk Factor C4A in Developmental Synaptic Pruning. Master's thesis, Harvard University. 2019. <https://doi.org/10.1038/s41593-020-00763-8>
167. Wangemann J. A human iPSC-based model for synaptic pruning-associated changes in schizophrenia. Doctoral dissertation. 2024.
168. Gracias J, Orhan F, Hörbeck E, Holmén-Larsson J, Khanlarkani N, Malwade S, Goparaju SK, Schwieler L, Demirel İŞ, Fu T, Fatourus-Bergman H. Cerebrospinal fluid concentration of complement component 4A is increased in first episode schizophrenia. *Nature Communications*. 2022 Nov 3;13(1):6427.
169. Prasad KM, Burgess AM, Keshavan MS, Nimgaonkar VL, Stanley JA. Neuropil pruning in early-course schizophrenia: immunological, clinical, and neurocognitive correlates. *Biol Psychiatry Cogn Neurosci Neuroimaging*. 2016;1(6):528-538.
170. Pearson CA, Iadecola C. When the BBB goes MIA. *Proc Natl Acad Sci U S A*. 2022;119(19):e2204159119.
171. Levkovitz Y, Mendlovich S, Riwkes S, Braw Y, Levkovitch-Verbin H, Gal G, et al. A double-blind, randomized study of minocycline for the treatment of negative and cognitive symptoms in early-phase schizophrenia. *J Clin Psychiatry*. 2010;71(2):138.
172. Oya K, Kishi T, Iwata N. Efficacy and tolerability of minocycline augmentation therapy in schizophrenia: a systematic review and meta-analysis of randomized controlled trials. *Hum Psychopharmacol Clin Exp*. 2014;29(5):483-491.
173. Solmi M, Veronese N, Thapa N, Facchini S, Stubbs B, Fornaro M, et al. Systematic review and meta-analysis of the efficacy and safety of minocycline in schizophrenia. *CNS Spectr*. 2017;22(5):415–26.
174. Deakin B, Suckling J, Barnes TR, Byrne K, Chaudhry IB, Dazzan P, et al. The benefit of minocycline on negative symptoms of schizophrenia in patients with recent-onset psychosis (BenEMin): a randomised, double-blind, placebo-controlled trial. *Lancet Psychiatry*. 2018;5(11):885–94.
175. Inta D, Lang UE, Borgwardt S, Meyer-Lindenberg A, Gass P. Microglia activation and schizophrenia: lessons from the effects of minocycline on postnatal neurogenesis, neuronal survival and synaptic pruning. *Schizophr Bull*. 2017;43(3):493–6.
176. De Picker LJ, Morrens M, Chance SA, Boche D. Microglia and brain plasticity in acute psychosis and schizophrenia illness course: A meta-review. *Front Psychiatry*. 2017;8:238.
177. Kato TA, Monji A, Mizoguchi Y, Hashioka S, Horikawa H, Seki Y, et al. Anti-inflammatory properties of antipsychotics via microglia modulations: Are antipsychotics a 'fire extinguisher' in the brain of schizophrenia? *Mini Rev Med Chem*. 2011;11(7):565–74.
178. Kroken RA, Sommer IE, Steen VM, Dieset I, Johnsen E. Constructing the immune signature of schizophrenia for clinical use and research: An integrative review translating descriptives into diagnostics. *Front Psychiatry*. 2019;9:753.
179. Bian Q, Kato T, Monji A, Hashioka S, Mizoguchi Y, Horikawa H, et al. The effect of atypical antipsychotics, perospirone, ziprasidone and quetiapine on microglial activation induced by interferon-gamma. *Prog Neuropsychopharmacol Biol Psychiatry*. 2008;32(1):42–8.
180. Hou Y, Wu CF, Yang JY, He X, Bi XL, Yu L, et al. Effects of clozapine, olanzapine and haloperidol on nitric oxide production by lipopolysaccharide-activated N9 cells. *Prog Neuropsychopharmacol Biol Psychiatry*. 2006;30:1523–8.

181. Kowalski J, Labuzek K, Herman ZS. Flupentixol and trifluoperidol reduce secretion of tumor necrosis factor- α and nitric oxide by rat microglial cells. *Neurochem Int.* 2003;43:173–8.
182. Kowalski J, Labuzek K, Herman ZS. Flupentixol and trifluoperidol reduce interleukin- 1β and interleukin-2 release by rat mixed glial and microglial cell cultures. *Pol J Pharmacol.* 2004;56:563–70.
183. Labuzek K, Kowalski J, Gabryel B, Herman ZS. Chlorpromazine and loxapine reduce interleukin- 1β and interleukin-2 release by rat mixed glial and microglial cell cultures. *Eur Neuropsychopharmacol.* 2005;15:23–30.
184. Coyle JT. Glutamate and schizophrenia: beyond the dopamine hypothesis. *Cell Mol Neurobiol.* 2006;26:365–84. doi:10.1007/s10571-006-9062-8. <https://doi.org/10.1007/s10571-006-9062-8>
185. Goff DC, Coyle JT. The emerging role of glutamate in the pathophysiology and treatment of schizophrenia. *American Journal of Psychiatry.* 2001 Sep 1;158(9):1367-77.
186. Moghaddam B, Javitt D. From revolution to evolution: the glutamate hypothesis of schizophrenia and its implication for treatment. *Neuropsychopharmacology.* 2012;37:4–15. doi: 10.1038/npp.2011.181.
187. Howes OD, Shatalina E. Integrating the neurodevelopmental and dopamine hypotheses of schizophrenia and the role of cortical excitation-inhibition balance. *Biol Psychiatry.* 2022;92(6):501–13.
188. Orhan F, Malwade S, Khanlarkhani N, Gkoga A, Jungholm O, Koskuvi M, Lehtonen Š, Schwieler L, Jardemark K, Tiihonen J, Koistinaho J. Kynurenic acid promotes activity-dependent synaptic pruning in schizophrenia. *bioRxiv.* 2023 Oct 23:2023-10.
189. Kasai H, Fukuda M, Watanabe S, Hayashi-Takagi A, Noguchi J. Structural dynamics of dendritic spines in memory and cognition. *Trends in neurosciences.* 2010 Mar 1;33(3):121-9.
190. Girdhar K, Rahman S, Dong P, Fullard JF, Roussos P. The neuroepigenome: implications of chemical and physical modifications of genomic DNA in schizophrenia. *Biological psychiatry.* 2022 Sep 15;92(6):443-9.
191. Gardoni F, Stanic J, Scheggia D, Benussi A, Borroni B, Di Luca M. NMDA and AMPA receptor autoantibodies in brain disorders: from molecular mechanisms to clinical features. *Cells.* 2021;10. <https://doi.org/10.3390/cells10010077>
192. Pollak TA, McCormack R, Peakman M, Nicholson TR, David AS. Prevalence of anti-N-methyl-D-aspartate (NMDA) receptor antibodies in patients with schizophrenia and related psychoses: a systematic review and meta-analysis. *Psychol Med.* 2014;44(12):2475-2487.
193. Hua JY, Smith SJ. Neural activity and the dynamics of central nervous system development. *Nature neuroscience.* 2004 Apr 1;7(4):327-32.
194. McKinney RA. Excitatory amino acid involvement in dendritic spine formation, maintenance and remodelling. *The Journal of physiology.* 2010 Jan 1;588(1):107-16.
195. Smith SS, Benanni S, Jones Q, Kenney L, Evrard M. Manipulation of $\alpha 4\beta\delta$ GABAA receptors alters synaptic pruning in layer 3 prelimbic prefrontal cortex and impairs temporal order recognition: Implications for schizophrenia and autism. *Brain Res.* 2024;148929.
196. Ghit A, Assal D, Al-Shami AS, Hussein DEE. GABAA receptors: structure, function, pharmacology, and related disorders. *J Genet Eng Biotechnol.* 2021;19(1):123.
197. Ge S, Goh ELK, Sailor KA, et al. GABA regulates synaptic integration of newly generated neurons in the adult brain. *Nature.* 2006;439:589–93.

198. Duveau V, Laustela S, Barth L, et al. Spatiotemporal specificity of GABAA receptor-mediated regulation of adult hippocampal neurogenesis. *Eur J Neurosci*. 2011;34:362–73.
199. Lee V, MacKenzie G, Hooper A, Maguire J. Reduced tonic inhibition in the dentate gyrus contributes to chronic stress-induced impairments in learning and memory. *Hippocampus*. 2016;26:1276–90.
200. Martin LJ, Zurek AA, MacDonald JF, et al. Alpha5GABAA receptor activity sets the threshold for long-term potentiation and constrains hippocampus-dependent memory. *J Neurosci*. 2010;30:5269–5282.
201. Schafer DP, Lehrman EK, Kautzman AG, Koyama R, Mardinly AR, Yamasaki R, et al. Microglia sculpt postnatal neural circuits in an activity and complement-dependent manner. *Neuron*. 2012;74(4):691–705.
202. Sipe GO, Lowery RL, Tremblay ME, Kelly EA, Lamantia CE, Majewska AK. Microglial P2Y12 is necessary for synaptic plasticity in mouse visual cortex. *Nat Commun*. 2016;7:10905.
203. Tremblay ME, Lowery RL, Majewska AK. Microglial interactions with synapses are modulated by visual experience. *PLoS Biol*. 2010;8:e1000527.
204. Plata ALD, Robles E. NMDA receptor antagonist MK801 reduces dendritic spine density and stability in zebrafish pyramidal neurons. *Neuroscience*. 2022;498:50-63.
205. Ultanir SK, Kim JE, Hall BJ, Deerinck T, Ellisman M, Ghosh A. Regulation of spine morphology and spine density by NMDA receptor signaling in vivo. *Proc Natl Acad Sci*. 2007;104(49):19553-19558.
206. Lewis DA, Moghaddam B. Cognitive dysfunction in schizophrenia: convergence of gamma-aminobutyric acid and glutamate alterations. *Arch Neurol*. 2006;63:1372–1376.
207. Malhotra AK, et al. Ketamine-induced exacerbation of psychotic symptoms and cognitive impairment in neuroleptic-free schizophrenics. *Neuropsychopharmacology*. 1997;17:141–150.
208. Jeevakumar V, et al. Ketamine administration during the second postnatal week induces enduring schizophrenia-like behavioral symptoms and reduces parvalbumin expression in the medial prefrontal cortex of adult mice. *Behav Brain Res*. 2015;282:165–75.
209. Martínez-Pinteño A, et al. The positive allosteric modulator of the mGlu2 receptor JNJ-46356479 partially improves neuropathological deficits and schizophrenia-like behaviors in a postnatal ketamine mice model. *J Psychiatr Res*. 2020;126:8–18.
210. Rabinovitch A, Braunstein D, Rabinovitch R, Biton Y. Possible mechanism of schizophrenia origin by excess GABA and synaptic pruning. *IBRO Neuroscience Reports*. 2023 Dec 1;15:126-30.
211. Tiihonen J, Wahlbeck K. Glutamatergic drugs for schizophrenia. *Cochrane Database of Systematic Reviews*. 2006(2).
212. Errico F, Mothet JP, Usiello A. D-Aspartate: An endogenous NMDA receptor agonist enriched in the developing brain with potential involvement in schizophrenia. *J Pharm Biomed Anal*. 2015;116:7–17.
213. Gos T, Myint AM, Schiltz K, Meyer-Lotz G, Dobrowolny H, Busse S, et al. Reduced microglial immunoreactivity for endogenous NMDA receptor agonist quinolinic acid in the hippocampus of schizophrenia patients. *Brain Behav Immun*. 2014;41:59–64.
214. Engel M, Snikeris P, Matosin N, Newell KA, Huang XF, Frank E. mGluR2/3 agonist LY379268 rescues NMDA and GABAA receptor level deficits induced in a two-hit mouse model of schizophrenia. *Psychopharmacology*. 2016;233:1349–59.

215. Errico F, Rossi S, Napolitano F, Catuogno V, Topo E, Fisone G, et al. D-aspartate prevents corticostriatal long-term depression and attenuates schizophrenia-like symptoms induced by amphetamine and MK-801. *J Neurosci*. 2008;28(41):10404–14.
216. Liebman HM, Woo W. THE ADDITION OF TIAGABINE TO ANTIPSYCHOTIC MEDICATION IN THE TREATMENT OF RECENT-ONSET SCHIZOPHRENIA BY MODIFICATION OF DEVELOPMENTAL PRUNING OF PREFRONTAL CIRCUITRY. *Schizophrenia Research*. 2010 Apr 1;117(2-3):380-1.
217. Burke, E., Wojcik, J., Seidman, L. J., Green, A., & Woo, T. U. W. (2018). F43. Potentiation of inhibitory neurotransmission in the treatment of recent-onset schizophrenia by modification of developmental pruning of prefrontal circuitry. *Schizophrenia Bulletin*.
218. Bosch C, Masachs N, Exposito-Alonso D, Martínez A, Teixeira CM, Fernaud I, et al. Reelin regulates the maturation of dendritic spines, synaptogenesis and glial ensheathment of newborn granule cells. *Cereb Cortex*. 2016;26:4282–98.
219. Niu S, Yabut O, D'Arcangelo G. The Reelin signaling pathway promotes dendritic spine development in hippocampal neurons. *Journal of Neuroscience*. 2008 Oct 8;28(41):10339-48.
220. Pujadas L, Gruart A, Bosch C, Delgado L, Teixeira CM, Rossi D, et al. Reelin regulates postnatal neurogenesis and enhances spine hypertrophy and long-term potentiation. *J Neurosci*. 2010;30:4636–4649.
221. Teixeira CM, Kron MM, Masachs N, Zhang H, Lagace DC, Martinez A, et al. Cell-autonomous inactivation of the Reelin pathway impairs adult neurogenesis in the hippocampus. *J Neurosci*. 2012;32:12051–12065.
222. González-Billault C, Del Río JA, Ureña JM, Jiménez-Mateos EM, Barallobre MJ, Pascual M, et al. A role of MAP1B in Reelin-dependent neuronal migration. *Cereb Cortex*. 2005;15:1134–45.
223. Molnár Z, Clowry GJ, Šestan N, Alzu'bi A, Bakken T, Hevner RF, et al. New insights into the development of the human cerebral cortex. *J Anat*. 2019;235:432–451.
224. Simó S, Jossin Y, Cooper JA. Cullin 5 regulates cortical layering by modulating the speed and duration of Dab1-dependent neuronal migration. *J Neurosci*. 2010;30:5668–76.
225. Simó S, Pujadas L, Segura MF, La Torre A, Del Río JA, Ureña JM, et al. Reelin induces the detachment of postnatal subventricular zone cells and the expression of the Egr-1 through Erk1/2 activation. *Cereb Cortex*. 2007;17:294–303.
226. Yasui N, Nogi T, Takagi J. Structural basis for specific recognition of Reelin by its receptors. *Structure*. 2010;18:320–331.
227. Walton E, Hass J, Liu J, Roffman JL, Bernardoni F, Roessner V, Kirsch M, Schackert G, Calhoun V, Ehrlich S. Correspondence of DNA methylation between blood and brain tissue and its application to schizophrenia research. *Schizophrenia bulletin*. 2016 Mar 1;42(2):406-14.
228. Impagnatiello F, Guidotti AR, Pesold C, Dwivedi Y, Caruncho H, Pisu MG, et al. A decrease of reelin expression as a putative vulnerability factor in schizophrenia. *Proc Natl Acad Sci USA*. 1998;95(26):15718–23
229. Yin J, Lu Y, Yu S, Dai Z, Zhang F, Yuan J. Exploring the mRNA expression level of RELN in peripheral blood of schizophrenia patients before and after antipsychotic treatment. *Hereditas*. 2020;157:1-6.

230. Fikri RM, Norlelawati AT, El-Huda AR, Hanisah MN, Kartini A, Norsidah K, Zamzila AN. Reelin (RELN) DNA methylation in the peripheral blood of schizophrenia. *Journal of psychiatric research*. 2017 May 1;88:28-37.
231. Markiewicz R, Markiewicz-Gospodarek A, Borowski B, Trubalski M, Łoza B. Reelin signaling and synaptic plasticity in schizophrenia. *Brain Sciences*. 2023 Dec 11;13(12):1704.
232. Boksa P. Abnormal synaptic pruning in schizophrenia: Urban myth or reality?. *Journal of Psychiatry and Neuroscience*. 2012 Mar 1;37(2):75-7.
233. Chai X, Förster E, Zhao S, Bock HH, Frotscher M. Reelin stabilizes the actin cytoskeleton of neuronal processes by inducing n-cofilin phosphorylation at serine3. *Journal of Neuroscience*. 2009 Jan 7;29(1):288-99.
234. Guidotti A, Grayson DR, Caruncho HJ. Epigenetic RELN dysfunction in schizophrenia and related neuropsychiatric disorders. *Front Cell Neurosci*. 2016;10:89.
235. Hill JJ, Hashimoto T, Lewis DA. Molecular mechanisms contributing to dendritic spine alterations in the prefrontal cortex of subjects with schizophrenia. *Molecular psychiatry*. 2006 Jun;11(6):557-66.
236. Hayashi-Takagi A, Takaki M, Graziane N, Seshadri S, Murdoch H, Dunlop AJ, Makino Y, Seshadri AJ, Ishizuka K, Srivastava DP, Xie Z. Disrupted-in-Schizophrenia 1 (DISC1) regulates spines of the glutamate synapse via Rac1. *Nature neuroscience*. 2010 Mar;13(3):327-32.
237. Lee FH, Kaidanovich-Beilin O, Roder JC, Woodgett JR, Wong AH. Genetic inactivation of GSK3 α rescues spine deficits in Disc1-L100P mutant mice. *Schizophrenia research*. 2011 Jun 1;129(1):74-9.
238. Roberts RC. Schizophrenia in translation: disrupted in schizophrenia (DISC1): integrating clinical and basic findings. *Schizophr Bull*. 2007;33:11–5.
239. Chandran JS, Kazanis I, Clapcote SJ, Ogawa F, Millar JK, Porteous DJ, et al. Disc1 variation leads to specific alterations in adult neurogenesis. *PLoS One*. 2014;9:e108088.
240. Nakata K, Lipska BK, Hyde TM, Ye T, Newburn EN, Morita Y, et al. DISC1 splice variants are upregulated in schizophrenia and associated with risk polymorphisms. *Proc Natl Acad Sci U S A*. 2009;106(37):15873–15878.
241. Rastogi A, Zai C, Likhodi O, Kennedy JL, Wong AH. Genetic association and post-mortem brain mRNA analysis of DISC1 and related genes in schizophrenia. *Schizophr Res*. 2009;114(1–3):39–49.
242. Schumacher J, Laje G, Abou Jamra R, Becker T, Muhleisen TW, Vasilescu C, et al. The DISC locus and schizophrenia: evidence from an association study in a central European sample and from a meta-analysis across different European populations. *Hum Mol Genet*. 2009;18(14):2719–27.
243. Xu Y, Ren J, Ye H. Association between variations in the disrupted in schizophrenia 1 gene and schizophrenia: A meta-analysis. *Gene*. 2018;651:94-99.
244. Dahoun T, Trossbach SV, Brandon NJ, Korth C, Howes OD. The impact of Disrupted-in-Schizophrenia 1 (DISC1) on the dopaminergic system: a systematic review. *Transl Psychiatry*. 2017;7(1):e1015.
245. Bellon A, Feuillet V, Cortez-Resendiz A, Mouaffak F, Kong L, Hong LE, De Godoy L, Jay TM, Hosmalin A, Krebs MO. Dopamine-induced pruning in monocyte-derived-neuronal-like cells (MDNCs) from patients with schizophrenia. *Molecular Psychiatry*. 2022 Jun;27(6):2787-802.
246. Cocchi E, Drago A, Serretti A. Hippocampal pruning as a new theory of schizophrenia etiopathogenesis. *Molecular neurobiology*. 2016 Apr;53(3):2065-81.

247. Bamne M, Wood J, Chowdari K, Watson AM, Celik C, Mansour H, et al. Evaluation of HLA polymorphisms in relation to schizophrenia risk and infectious exposure. *Schizophr Bull.* 2012;38(6):1149–54.
248. Purcell SM, et al. Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. *Nature.* 2009;460:748–752.
249. Shi J, Levinson DF, Duan J, Sanders AR, Zheng Y, Pe'er I, et al. Common variants on chromosome 6p22.1 are associated with schizophrenia. *Nature.* 2009;460:753–7.
250. Stefansson H, et al. Common variants conferring risk of schizophrenia. *Nature.* 2009;460:744–747.
251. Brucato N, Guadalupe T, Franke B, Fisher SE, Francks C. A schizophrenia-associated HLA locus affects thalamus volume and asymmetry. *Brain Behav Immun.* 2015;46:311–8.
252. Rao NP, Kalmady S, Arasappa R, Venkatasubramanian G. Clinical correlates of thalamus volume deficits in antipsychotic-naive schizophrenia patients: a 3-Tesla MRI study. *Indian J Psychiatry.* 2010;52:229-235.
253. Knochel C, Stablein M, Prvulovic D, Ghinea D, Wenzler S, Pantel J, et al. Shared and distinct gray matter abnormalities in schizophrenia, schizophrenia relatives and bipolar disorder in association with cognitive impairment. *Schizophr Res.* 2016.
254. Pathania M, Davenport EC, Muir J, Sheehan DF, López-Doménech G, Kittler JT. The autism and schizophrenia associated gene CYFIP1 is critical for the maintenance of dendritic complexity and the stabilization of mature spines. *Transl Psychiatry.* 2014;4(3):e374.
255. Drew J, Arancibia-Carcamo IL, Jolivet R, Lopez-Domenech G, Attwell D, Kittler JT. Control of microglial dynamics by Arp2/3 and the autism and schizophrenia-associated protein Cyfip1. *bioRxiv.* 2020. <https://doi.org/10.1101/2020.05>.
256. Cioni JM, Wong HH-W, Bressan D, Kodama L, Harris WA, Holt CE. Axon-Axon Interactions Regulate Topographic Optic Tract Sorting via CYFIP2-Dependent WAVE Complex Function. *Neuron.* 2018;97:1078–93.e6.
257. Davenport EC, Szulc BR, Drew J, Taylor J, Morgan T, Higgs NF, et al. Autism and schizophrenia-associated CYFIP1 regulates the balance of synaptic excitation and inhibition. *Cell Rep.* 2019;26(8):2037–51.
258. Domínguez-Iturza N, Lo AC, Shah D, Armendáriz M, Vannelli A, Mercaldo V, et al. The autism- and schizophrenia-associated protein CYFIP1 regulates bilateral brain connectivity and behaviour. *Nat Commun.* 2019;10:3454.
259. De Rubeis S, Pasciuto E, Li KW, Fernández E, Di Marino D, Buzzi A, et al. CYFIP1 coordinates mRNA translation and cytoskeleton remodeling to ensure proper dendritic spine formation. *Neuron.* 2013;79:1169–82.
260. Hsiao K, Harony-Nicolas H, Buxbaum JD, Bozdagi-Gunal O, Benson DL. Cyfip1 regulates presynaptic activity during development. *J Neurosci.* 2016;36(6):1564–76.
261. Selemon LD, Goldman-Rakic PS. The reduced neuropil hypothesis: a circuit based model of schizophrenia. *Biological psychiatry.* 1999 Jan 1;45(1):17-25.
262. Selemon LD, Rajkowska G. Cellular pathology in the dorsolateral prefrontal cortex distinguishes schizophrenia from bipolar disorder. *Current Molecular Medicine.* 2003 Aug 1;3(5):427-36.
263. McGlashan TH, Hoffman RE. Schizophrenia as a disorder of developmentally reduced synaptic connectivity. *Archives of general psychiatry.* 2000 Jul 1;57(7):637-48.

264. Ducharme G, Lowe GC, Goutagny R, Williams S. Early alterations in hippocampal circuitry and theta rhythm generation in a mouse model of prenatal infection: implications for schizophrenia. *PLoS One*. 2012;7:e29754. <https://doi.org/10.1371/journal.pone.0029754>
265. Kühn S, Musso F, Mobascher A, et al. Hippocampal subfields predict positive symptoms in schizophrenia: first evidence from brain morphometry. *Transl Psychiatry*. 2012;2:e127.
266. Selten JP, Van Der Ven E, Rutten BP, Cantor-Graae E. The social defeat hypothesis of schizophrenia: an update. *Schizophr Bull*. 2013;39(6):1180–6.
267. Sheridan MA, McLaughlin KA. Dimensions of early experience and neural development: deprivation and threat. *Trends Cogn Sci*. 2014;18(11):580–85.
268. McLaughlin KA, Sheridan MA, Nelson CA. Neglect as a violation of species-expectant experience: neurodevelopmental consequences. *Biol Psychiatry*. 2017;82(7):462-471.
269. Piccolo LR, Merz EC, He X, Sowell ER, Noble KG, Pediatric Imaging, Neurocognition, Genetics Study. Age-related differences in cortical thickness vary by socioeconomic status. *PLoS One*. 2016;11(9):e0162511.
270. Malave L, van Dijk MT, Anacker C. Early life adversity shapes neural circuit function during sensitive postnatal developmental periods. *Transl Psychiatry*. 2022;12(1):306.
271. Zetter MA, Hernández VS, Roque A, Hernández-Pérez OR, Gómora MJ, Ruiz-Velasco S, et al. Microglial synaptic pruning on axon initial segment spines of dentate granule cells: Sexually dimorphic effects of early-life stress and consequences for adult fear response. *J Neuroendocrinol*. 2021;33(7):e12969.
272. Dayananda KK, Ahmed S, Wang D, Polis B, Islam R, Kaffman A. Early life stress impairs synaptic pruning in the developing hippocampus. *Brain Behav Immun*. 2023;107:16–31.
273. Hanson JL, van den Bos W, Roeber BJ, Rudolph KD, Davidson RJ, Pollak SD. Early adversity and learning: implications for typical and atypical behavioral development. *J Child Psychol Psychiatry*. 2017;58(7):770–8.
274. Wang J, Chen HS, Li HH, Wang HJ, Zou RS, Lu XJ, et al. Microglia-dependent excessive synaptic pruning leads to cortical underconnectivity and behavioral abnormality following chronic social defeat stress in mice. *Brain Behav Immun*. 2023;109:23-36.
275. Diwadkar VA, Flaugh B, Jones T, Zalányi L, Ujfalussy B, Keshavan MS, Erdi P. Impaired associative learning in schizophrenia: behavioral and computational studies. *Cogn Neurodyn*. 2008;2(3):207–19.
276. Reddaway J. The role of the complement system in associative learning and risk for schizophrenia [dissertation]. Cardiff University; 2022.
277. Smith KE, Pollak SD. Early life stress and development: potential mechanisms for adverse outcomes. *J Neurodev Disord*. 2020;12:34.
278. Hartley CA, Lee FS. Sensitive periods in affective development: nonlinear maturation of fear learning. *Neuropsychopharmacology*. 2015;40:50–60.
279. Reh RK, Dias BG, Nelson CA, Kaufer D, Werker JF, Kolb B, et al. Critical period regulation across multiple timescales. *Proc Natl Acad Sci U S A*. 2020;117:23242–23251.
280. Goodwill HL, Manzano-Nieves G, LaChance P, Teramoto S, Lin S, Lopez C, et al. Early life stress drives sex-selective impairment in reversal learning by affecting parvalbumin interneurons in orbitofrontal cortex of mice. *Cell Rep*. 2018;25:2299–307.

281. Murthy S, Kane GA, Katchur NJ, Lara Mejia PS, Obiofuma G, Buschman TJ, et al. Perineuronal nets, inhibitory interneurons, and anxiety-related ventral hippocampal neuronal oscillations are altered by early life adversity. *Biol Psychiatry*. 2019;85:1011–1020.
282. Kaar SJ, Angelescu I, Marques TR, Howes OD. Pre-frontal parvalbumin interneurons in schizophrenia: a meta-analysis of post-mortem studies. *J Neural Transm*. 2019;126:1637–51.
283. Schmitt A, Falkai P, Papiol S. Neurodevelopmental disturbances in schizophrenia: evidence from genetic and environmental factors. *J Neural Transm*. 2023;130(3):195–205.
284. Wekerle H. Planting and pruning in the brain: MHC antigens involved in synaptic plasticity? *Proc Natl Acad Sci*. 2005;102(1):3-4.
285. Hui CW, St-Pierre A, El Hajj H, Remy Y, Hébert SS, Luheshi GN, et al. Prenatal immune challenge in mice leads to partly sex-dependent behavioral, microglial, and molecular abnormalities associated with schizophrenia. *Front Mol Neurosci*. 2018;11:13.
286. Cattane N, Richetto J, Cattaneo A. Prenatal exposure to environmental insults and enhanced risk of developing schizophrenia and autism spectrum disorder: focus on biological pathways and epigenetic mechanisms. *Neurosci Biobehav Rev*. 2020;117:253–78.
287. Dietz AG, Goldman SA, Nedergaard M. Glial cells in schizophrenia: a unified hypothesis. *Lancet Psychiatry*. 2019;7(3):272–81.
288. Gumusoglu SB, Stevens HE. Maternal inflammation and neurodevelopmental programming: a review of preclinical outcomes and implications for translational psychiatry. *Biol Psychiatry*. 2019;85(2):107–21.
289. Howes OD, McCutcheon R. Inflammation and the neural diathesis-stress hypothesis of schizophrenia: a reconceptualization. *Transl Psychiatry*. 2017;7(2):e1024.
290. Davis J, Moylan S, Harvey BH, Maes M, Berk M. Neuroprogression in schizophrenia: pathways underpinning clinical staging and therapeutic corollaries. *Aust N Z J Psychiatry*. 2014;48(6):512–29.
291. Misiak B, Stramecki F, Gawęda Ł, Prochwicz K, Szaśniadek MM, Moustafa AA, et al. Interactions between variation in candidate genes and environmental factors in the etiology of schizophrenia and bipolar disorder: a systematic review. *Mol Neurobiol*. 2018;55:5075-5100.
292. Rao JS, Kellom M, Kim HW, Rapoport SI, Reese EA. Neuroinflammation and synaptic loss. *Neurochem Res*. 2012;37:903–910.
293. Hinwood M, Kluge MG, Ilicic M, Walker FR. Understanding microglial involvement in stress-induced mood disturbance: a modulator of vulnerability?. *Curr Opin Behav Sci*. 2019;28:98–104.
294. Weber MD, McKim DB, Niraula A, Witcher KG, Yin W, Sobol CG, et al. The influence of microglial elimination and repopulation on stress sensitization induced by repeated social defeat. *Biol Psychiatry*. 2019;85(8):667-678.
295. Montagud-Romero S, Montesinos J, Pavón FJ, Blanco-Gandia MC, Ballestín R, de Fonseca FR, et al. Social defeat-induced increase in the conditioned rewarding effects of cocaine: role of CX3CL1. *Prog Neuropsychopharmacol Biol Psychiatry*. 2020;96:109753.
296. Kato TA, Hayakawa K, Monji A, Kanba S. Missing and possible link between neuroendocrine factors, neuropsychiatric disorders, and microglia. *Front Integr Neurosci*. 2013;7:53.
297. Sugama S, Kakinuma Y. Stress and brain immunity: microglial homeostasis through hypothalamus-pituitary-adrenal gland axis and sympathetic nervous system. *Brain Behav Immun Health*. 2020;7:100111.

298. Sequeira MK, Bolton JL. Stressed Microglia: neuroendocrine–neuroimmune interactions in the stress response. *Endocrinology*. 2023;164(7):bqad088.
299. Cheiran Pereira G, Piton E, Moreira dos Santos B, Ramanzini LG, Muniz Camargo LF, Menezes da Silva R, et al. Microglia and HPA axis in depression: An overview of participation and relationship. *World J Biol Psychiatry*. 2022;23(3):165–82.
300. Bergon A, Belzeaux R, Comte M, Pelletier F, Herve M, Gardiner EJ, et al. CX3CR1 is dysregulated in blood and brain from schizophrenia patients. *Schizophr Res*. 2015;168(1-2):434–43.
301. Eack SM, Hogarty GE, Cho RY, Prasad KM, Greenwald DP, Hogarty SS, Keshavan MS. Neuroprotective effects of cognitive enhancement therapy against gray matter loss in early schizophrenia: results from a 2-year randomized controlled trial. *Arch Gen Psychiatry*. 2010;67(7):674–82.
302. Morimoto T, Matsuda Y, Matsuoka K, Yasuno F, Ikebuchi E, Kameda H, et al. Computer-assisted cognitive remediation therapy increases hippocampal volume in patients with schizophrenia: a randomized controlled trial. *BMC Psychiatry*. 2018;18:1-8.
303. Özdemir AA, Kavak Budak F. The effects of mindfulness-based stress reduction training on hope, psychological well-being, and functional recovery in patients with schizophrenia. *Clin Nurs Res*. 2022;31(2):183-193.
304. Halverson TF, Meyer-Kalos PS, Perkins DO, Gaylord SA, Palsson OS, Nye L, et al. Enhancing stress reactivity and wellbeing in early schizophrenia: a randomized controlled trial of Integrated Coping Awareness Therapy (I-CAT). *Schizophr Res*. 2021;235:91–101.
305. Steel C, Hardy A, Smith B, Wykes T, Rose S, Enright S, et al. Cognitive–behaviour therapy for post-traumatic stress in schizophrenia: A randomized controlled trial. *Psychol Med*. 2017;47(1):43-51.
306. Kicher T, Krug A, Cabanis M, et al. Neural correlates of cognitive behavioural therapy in patients with schizophrenia. *Eur Psychiatry*. 2011;26(S2):2156.